

Mitochondria and Tumorigenesis

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Abstract

Mitochondria are essential organelles in eukaryotes, serving as the cellular powerhouse for energy production. Mitochondrial dysregulation plays a pivotal role in disease progression by functioning as a "plasticity hub" that coordinates the dynamic tumor microenvironment. This coordination occurs via multidimensional mechanisms such as metabolic rewiring, bioenergetic flux disruption, stress-responsive signaling, and inter-organelle communication, collectively promoting cancer initiation, dissemination, and therapeutic evasion. The current review summarizes recent advances in understanding mitochondria-driven oncogenesis through these mechanisms, discusses mitochondrial-targeted therapeutic strategies and associated challenges, and aims to establish a cross-scale framework for decoding the multidimensional synergy of mitochondrial networks in cancer progression, thereby offering insights for developing innovative precision therapies that integrate metabolic-dynamic-immune interventions.

1. Mitochondrial Biology and Functions

As critical double-membraned eukaryotic organelles, mitochondria contain circular Mitochondrial DNA (mtDNA) that encodes 37 genes essential for oxidative phosphorylation - comprising polypeptides, tRNAs, and rRNAs [1,2]. Beyond their canonical role in bioenergy generation through oxidative phosphorylation (OXPHOS) and the tricarboxylic acid (TCA) cycle-mediated ATP production [3], mitochondria functionally extend to metabolic regulation, biosynthetic precursor generation, Ca²⁺ flux coordination, programmed cell death execution, and cellular signaling modulation (Figure 1) [4,5]. mtDNA mutations demonstrate oncogenic potential [6], particularly in malignancies such as breast [7], ovarian [7], and prostate cancers [8].

Cancer, a complex group of diseases, remains a major global health threat with rising incidence driven by population aging [9]. While molecularly-targeted agents and immune checkpoint inhibitors have revolutionized oncology, persistent obstacles including intratumoral heterogeneity, modality-specific constraints, and treatment refractoriness continue to impede clinical success. Consequently, the development of resilient treatment strategies represents an urgent unmet need. This work aims to establish a cross-scale framework elucidating mitochondria-driven multidimensional networks in cancer progression, while exploring innovative therapeutic strategies derived from these insights.

Functioning as a metabolic and signaling nexus, mitochondria establish multidimensional networks through metabolic plasticity, dynamic remodeling, epigenetic regulation, and immune microenvironment

reprogramming, acting as an "adaptive engine" that drives malignant transformation, metastasis, and therapeutic resistance (Figure 1). Nevertheless, therapeutic development faces substantial barriers, with mitochondrial-directed compounds frequently demonstrating constrained clinical utility owing to pathway redundancy and tumor cell-intrinsic adaptive mechanisms [10].

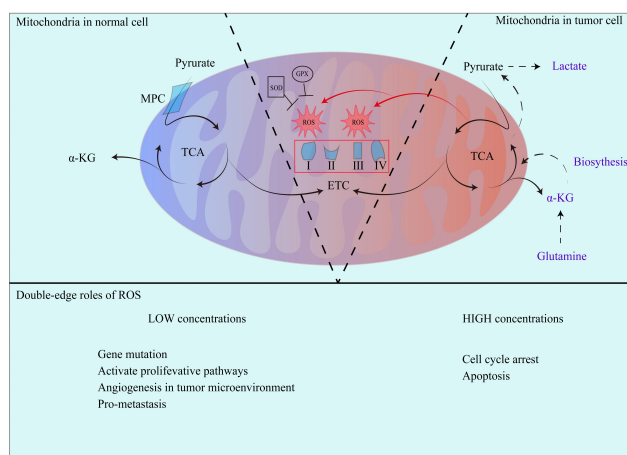


Figure 1. Mitochondrial Metabolic biology in Normal or Cancer Cell.

2. Mitochondrial Metabolic Reprogramming and Cancer

Mitochondrial metabolic reprogramming, characterized by profound functional alterations in cancer, serves as a central driver of malignant progression. By establishing dynamic, collaborative metabolic networks that transcend conventional energy metabolism frameworks,

it achieves multidimensional integration of energy production, redox regulation, and biosynthetic support - mechanisms fundamentally linked to carcinogenesis, metastasis, and therapeutic resistance [11,12]. Distinct from the metabolic homeostasis of normal cells, cancer cells develop spatiotemporally regulated metabolic plasticity through dynamic pathway switching and metabolite functional expansion, forming an adaptable metabolic ecosystem. This system not only fulfills the bioenergetic demands of rapid proliferation but also remodels the microenvironment, modulates immune responses, and drives metastasis/drug resistance through metabolic intermediates, ultimately securing survival advantages in heterogeneous microenvironments. Deciphering the functional modularity and interaction mechanisms within metabolic networks will provide novel perspectives for targeting metabolic vulnerabilities and overcoming therapeutic resistance.

2.1 Dynamic Equilibrium of Energy Metabolism

The malignant proliferation of tumor cells relies on dynamic remodeling of energy supply patterns. The Warburg effect describes cancer cells' preferential utilization of glycolysis over oxidative phosphorylation (OXPHOS) for ATP production through glucose fermentation to lactate, even under aerobic conditions with functional mitochondria [13]. Modern studies reveal the Warburg effect arises not from defective mitochondrial respiration as originally proposed [14], but rather serves tumor-specific biological requirements distinct from normal tissue physiology. Unlike normal cells' OXPHOS-dominated homeostasis, tumors develop unique metabolic plasticity through spatiotemporal coordination of Warburg effect and OXPHOS: glycolytic bursts support rapid proliferation, while mitochondrial respiration sustains metastasis and therapy resistance.

Emerging research demonstrates a sophisticated dynamic equilibrium between Warburg metabolism and OXPHOS, transcending simplistic binary opposition. While Kukurugya's metabolic model confirmed glycolysis's kinetic advantage in ATP production [13], glycolytic tumors paradoxically require mitochondrial functions. For instance, Metastasis-Associated Protein 1 (MTA1) enhances OXPHOS via ATP synthase upregulation to drive colorectal liver metastases [15], and prostate cancer cells shift from glycolysis to OXPHOS under high-density conditions [16], challenging the "glycolytic metastasis" dogma.

The classical Warburg paradigm is being redefined, as tumors exhibit metabolic flexibility to adapt their energy strategies according to microenvironmental constraints. Under proliferative or hypoxic stress, tumors activate glycolysis through mechanisms like OMA1-mediated metabolic coordination in colorectal cancer [17], while Skp2 maintains cell cycle-coupled metabolism - Tricarboxylic Acid (TCA) cycle in G1 phase and glycolysis in S phase via IDH1/2 stabilization in prostate cancer [18]. Conversely, glycolytic toxicity (e.g., lactic acidosis) induces OXPHOS revival, creating therapeutic vulnerabilities - tumor sensitivity to OXPHOS inhibitors increases 100-10,000 fold under these conditions [19].

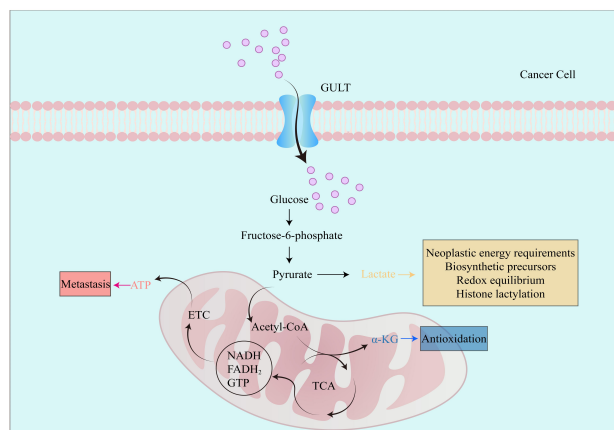


Figure 2. Metabolic-Epigenetic Nexus in Mitochondria of Cancer Cell: Glycolytic Reprogramming, Lactate-Driven Histone Modifications, and Redox Adaptation in Metastatic Progression.

2.2 Regulation of Redox Homeostasis

Under dual pressures of hypermetabolism and hostile microenvironments, cancer cells construct multi-layered redox defense networks through functional expansion of metabolic intermediates. Glutamine metabolism, TCA cycle reprogramming, and glycolytic byproducts form coordinated regulatory axes to combat oxidative stress. Notably, these metabolites not only maintain tumor-intrinsic redox balance but also remodel the microenvironment through epigenetic modifications and immune cell polarization, creating pro-tumorigenic ecological niches. Metabolic regulation of redox homeostasis unveils the profound evolutionary mechanism through which tumors convert survival stress into progression momentum.

In cancers, enhanced dependency on glutamine uptake and catabolism profoundly impacts redox regulation through metabolic reprogramming. Studies reveal glutamine "addiction" in various malignancies - tumor cells critically depend on glutamine as both energy substrate and biosynthetic precursor [20]. PDAC cells utilize an unconventional glutamine pathway: glutamine-derived aspartate undergoes cytosolic transport and GOT1-mediated conversion to oxaloacetate, then malate/pyruvate, elevating NADPH/NADP⁺ ratio to combat oxidative stress [21]. TCA cycle remodeling demonstrates environmental adaptability - α -ketoglutarate (KG) acts as antioxidant under oxidative stress, reducing Reactive Oxygen Species (ROS) production [22], enabling metabolic equilibrium maintenance in adverse conditions. The Warburg effect further regulates glutamine metabolism and ROS dynamics, enabling persistent redox homeostasis [23].

2.3 Biosynthetic Support

The ultimate objective of metabolic reprogramming is to supply biosynthetic precursors for uncontrolled tumor proliferation. By hijacking glycolytic branches, remodeling TCA cycle flux, and enhancing glutaminolysis, cancer cells redirect metabolic flows toward nucleotide, amino acid, and lipid precursor synthesis. For instance: enhanced glycolysis provides

abundant precursors for nucleotide/amino acid/lipid biosynthesis to sustain rapid division [24]; hypoxic melanoma cells utilize reverse TCA flux to synthesize fatty acids from glutamine, critical for proliferation [25]; glutaminolysis supplies nitrogen for nucleic acid synthesis, with increased glutamine uptake boosting ATP and macromolecule production [26]. PDAC cells employ a non-canonical glutamine pathway where aspartate-malate flux supports biosynthesis [21]. This "metabolic diversion" strategy not only surpasses normal anabolic rate limitations but also exploits non-canonical enzyme functions: glycolytic enzyme HK2 phosphorylates I κ B α to activate NF- κ B pathway, upregulating PD-L1 and directly mediating immune evasion [27], highlighting the central role of metabolic networks in malignant transformation.

2.4 Lipid Metabolic Reprogramming

Lipids (FAs, cholesterol, phospholipids, lipoproteins) fulfill essential physiological roles in membrane architecture, energy storage, and signal transduction, while playing critical roles in tumorigenesis, progression, and metastasis [28,29]. Lipid metabolic remodeling represents a critical evolutionary adaptation to energy crises and membrane biogenesis demands in malignancies. Upregulation of lipogenic regulators in cancers involves both aberrant de novo lipogenesis and enhanced exogenous lipid uptake. Amplified fatty acid synthesis provides structural membrane components and bioenergetic substrates to fuel rapid proliferation [30-32]. The unique lipid composition of cancer cell membranes is vital for survival, with tumors exhibiting both quantitative lipid accumulation and qualitative compositional changes to meet specialized physiological demands [33]. Alterations in lipid metabolism remodel the entire tumor microenvironment, impairing immune cell function and antitumor responses [34]. Tumor-specific lipid metabolic patterns demonstrate metabolic plasticity breadth while providing rationale for targeting lipogenic enzymes in combination with immunotherapy.

3. Mitochondrial Dynamics and Cancer Invasion

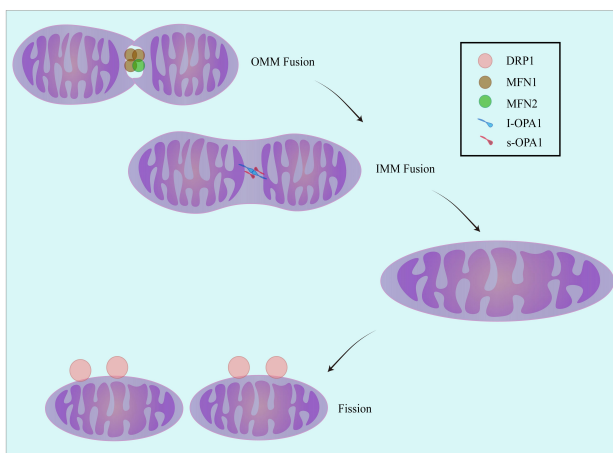


Figure 3. Mitochondrial Fusion and Fission.

Mitochondrial dynamics refers to the homeostatic process maintaining tubular networks through continuous

fusion/fission cycles (Figure 3), which orchestrates morphology, size, quantity, and distribution - essential for cellular physiology [35-37]. Perturbations in mitochondrial dynamics disrupt functional integrity, driving pathological progression including carcinogenesis [36], with emerging evidence highlighting its mechanistic link to metastatic dissemination [38,39].

3.1 Mitochondrial Fission-Driven Invasive Phenotypes

Mitochondrial fission, a tightly regulated process mediated by dynamin-related protein 1 (Drp1), governs organellar morphology and quantity while coordinating metabolic signaling and cell fate determination - mechanisms deeply implicated in oncogenesis [40-42]. Drp1-mediated fission promotes tumor progression, as evidenced in esophageal squamous cell carcinoma (ESCC) where Drp1 overexpression activates cGAS-STING-induced autophagy to drive malignancy [43]. Drp1 enhances metastatic competency across malignancies: metastatic breast cancers exhibit elevated Drp1 expression/activity [38]; lung adenocarcinoma migration correlates with Drp1 levels [41]; glioma invasion depends on Drp1-RHOA/ROCK1-mediated cytoskeletal remodeling via filopodia regulation [44]. Functional validation in triple-negative breast cancer (TNBC) shows pharmacogenetic induction of mitochondrial fragmentation paradoxically suppresses metastasis, challenging conventional paradigms [45].

Mitochondrial fragmentation via Drp1 facilitates Warburg metabolism, providing energetic/anabolic support for metastatic dissemination. Paradoxically, mitochondrial fission factor (MFF) amplification diminishes bioenergetic capacity through dual suppression of oxidative and glycolytic metabolism [46]. While fission generally promotes invasion, MFF-driven fragmentation unexpectedly reduces CSC viability, highlighting context-dependent metabolic regulation [46]. Latent brain metastases employ mitochondrial fragmentation to activate FAO, maintaining metabolic flexibility during microenvironmental adaptation [47]. Hepatocellular carcinoma models show fission activation upregulates lipogenesis while suppressing β -oxidation, redirecting glycolytic intermediates to support proliferation [48]. Fission remodels tumor-stroma crosstalk - prostate cancer CAFs engage in metabolic symbiosis with cancer cells [49], while PI3K/Akt/mTOR and MAPK pathways integrate fission signals to drive invasion [45].

3.2 Mitochondrial Fusion Deficits and Metastatic Potential

Mitochondrial fusion requires coordinated outer/inner membrane remodeling: outer membrane GTPases Mitofusins (Mfn1/2) mediate homologous interactions through complex formation [50-52], while inner membrane cristae maintenance involves OPA1 GTPase activity that correlates with OXPHOS efficiency [51,53]. Fusion homeostasis optimizes bioenergetic output, enables inter-mitochondrial mtDNA exchange [54], and

maintains functional integration with cellular physiology. Metastatic outcomes depend on fusion equilibrium - physiological fusion mitigates ROS toxicity [38], whereas excessive fusion triggers bioenergetic overload and mitotic catastrophe [55,56].

3.3 Imbalance of mitochondrial dynamics and cancer development

In summary, enhanced mitochondrial fission critically supports cancer cell proliferation/survival and drives metastasis via metabolic reprogramming, while fusion enhances metabolic adaptability and bioenergetic output to promote therapeutic resistance. Paradoxically, emerging evidence suggests fusion may exert tumor-suppressive effects in specific contexts. PDAC studies demonstrate that mitochondrial fusion restoration rescues cristae architecture, attenuates oxidative metabolism, and confers therapeutic vulnerability [57]. Genetic/pharmacological targeting of OPA1-mediated fusion in TNBC models impairs metastatic competency through bioenergetic crisis induction [58].

While fission and fusion exhibit context-dependent effects on cancer survival, malignant cells dynamically balance these processes to meet evolving metabolic and bioenergetic demands. Although core regulators (e.g., Drp1, Mfns) have been identified, their cancer-type-specific mechanisms and crosstalk with signaling pathways require deeper characterization, as do the metabolic heterogeneity and context-dependent vulnerabilities across malignancies.

4. Mitochondrial Stress Signaling

Mitochondrial stress signaling encompasses molecular cascades initiated under organellar duress, which propagate intracellularly to elicit adaptive responses [59,60]. In oncology, mitochondrial stress signaling has emerged as critical for tumor survival under microenvironmental stresses (hypoxia, nutrient scarcity), driving metabolic rewiring to sustain proliferation (Figure 4) [61-63].

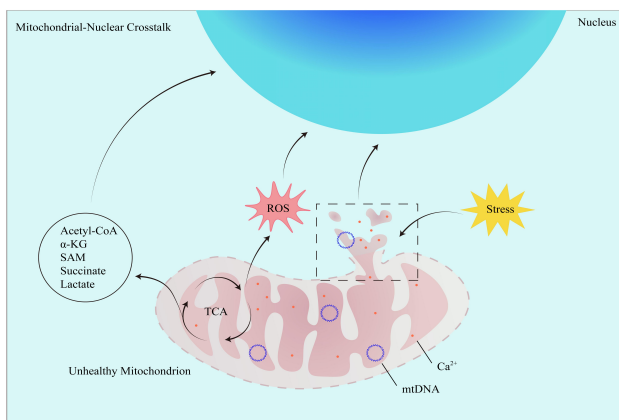


Figure 4. Mitochondrial Dysfunction Orchestrates Nuclear Reprogramming in Cancer Cells.

4.1 The Double-Edged Sword of ROS

Reactive oxygen species (ROS), highly reactive molecules generated through mitochondrial electron transport chain activity, serve as signaling mediators under physiological conditions, dynamically regulating tumor survival, metastasis, and therapy resistance (Figure 1). However, ROS imbalance induces oxidative stress, causing macromolecular damage while paradoxically modulating malignant progression [64-66].

At low concentrations, ROS act as pro-tumorigenic signals through DNA oxidation (e.g., guanine→8-oxodG [67]) and strand breaks. Remarkably, even subtoxic H₂O₂ levels induce oxidative clustered DNA lesions (OCDLs) and replication-independent DSBs [67]. ROS activate proliferative pathways like Ras-Raf-MEK-ERK cascade [68,69], where sustained ERK1/2 phosphorylation drives G1/S transition through cyclin induction and tumor suppressor silencing [70,71]. Vascular Endothelial Growth Factor (VEGF) plays a central role in tumor angiogenesis [72], which can be upregulated by reactive oxygen species (ROS) through stabilizing and activating hypoxia-inducible factor 1 α (HIF-1 α) and activating the nuclear factor κ B (NF- κ B) pathway, while its expression is also induced by MAPK pathway upregulation [73-76]. Additionally, ROS activate EMT-associated transcription factors to induce epithelial-mesenchymal transition in tumor cells, enhancing invasion and metastasis [77,78]. ROS also remodel tumor microenvironments by inducing immune cell apoptosis/functional suppression [79,80] and promoting normal fibroblast transdifferentiation into cancer-associated fibroblasts (CAFs) [81,82], thereby creating a protumorigenic niche.

Supraphysiological ROS activate counter-regulatory pathways like p38 MAPK, inducing G2/M arrest by antagonizing pro-growth ERK signaling [68,83-85]. Excessive ROS activate both intrinsic and extrinsic apoptotic pathways: 1) ROS oxidize mitochondrial membrane components (cardiolipin/VDAC), increasing permeability and cytochrome c release [86-89]; 2) ROS upregulate pro-apoptotic Bax while downregulating anti-apoptotic Bcl-2/Mcl-1, enhancing MOMP [90-92]; 3) Cytochrome c forms apoptosomes with Apaf-1 and procaspase-9, activating caspase-9/-3 cascade [93,94]. In the extrinsic pathway, ROS: 1) Upregulate death receptors (Fas/DR4/DR5) [95]; 2) Enhance FasL-receptor binding [96]; 3) Activate caspase-8 via FADD adaptors, initiating effector caspase cascade [97]. ROS induce ER stress-mediated apoptosis via: 1) Calcium dysregulation through oxidized channels [98,99]; 2) PERK-eIF2 α -ATF4 axis activation causing CHOP overexpression [100-102]; 3) Caspase-12 activation initiating executioner caspases [103,104]. Paradoxically, while ROS-induced DNA damage drives mutagenesis, catastrophic genomic lesions ultimately activate apoptotic checkpoints [105].

4.2 Mitochondrial Unfolded Protein Response (UPR^{mt})

UPR^{mt} represents an evolutionarily conserved adaptive mechanism that preserves mitochondrial proteostasis by upregulating chaperones and proteases through stress-responsive transcriptional programs. Proteotoxic stress induces UPR^{mt}-mediated transcriptional adaptation, deploying HSP60/LONP1 complexes to eliminate damaged proteins [106-108]. UPR^{mt} activation initiates adaptive programs facilitating mitochondrial recovery, metabolic plasticity, and innate immune modulation [109]. Cancer cells hijack UPR^{mt} to resolve replication stress-induced proteotoxicity, maintaining mitochondrial fitness during malignant progression [110,111]. The UPR^{mt} maintains mitochondrial proteostasis to support pre-metastatic adaptation, and targeting its critical regulatory nodes could enhance chemotherapeutic sensitivity.

Neoplastic mitochondria endure dual assaults: electron transport chain-derived oxidative damage and replication error-prone mtDNA synthesis. Unchecked mtROS induces proteotoxic collapse, necessitating UPR^{mt}-regulated chaperone-mediated refolding and protease-dependent degradation of damaged proteins [110]. Malignant subpopulations exhibit UPR^{mt} hyperactivation as pre-adaptive strategy, maintaining redox equilibrium to fuel metastatic outgrowth [112]. Mitochondrial proteotoxicity induces: 1) ATF5 nuclear translocation activating UPR^{mt} genes [113]; 2) ROS-mediated DNAJA1 oxidation enhancing HSP70-c-mtProt interaction [114]; 3) HSF1 liberation driving compensatory transcriptional programs [114].

Non-pathological UPR^{mt} induction confers systemic benefits via mitohormetic strengthening of stress response networks [112]. Oncogenic UPR^{mt} reprogramming creates self-reinforcing loops that sustain proliferative and metastatic programs [111]. In prostate cancer, the HSP60-ClpP axis sustains malignancy through c-Myc-driven expression and functional coordination, restoring oncogenic mitochondrial activity [115]. The SIRT3-UPR^{mt} axis drives metastasis, with high UPR^{mt} activity signatures (7-gene panel) predicting poor outcomes in breast cancer patients [112]. UPR^{mt}-upregulated antioxidants (SOD1/2) establish "mitochondrial fitness zones" that neutralize oxidative insults in tumors [116].

5. Mitochondrial-Nuclear Crosstalk

Mitochondria maintain residual genomes but predominantly rely on nuclear-encoded proteins imported from cytosol. This genomic segregation necessitates sophisticated mitochondrial-nuclear communication to preserve cellular functionality and adaptability [117-119]. This inter-organellar dialogue integrates metabolic reprogramming, death signaling, ionic balance, and adaptive survival mechanisms under pathophysiological conditions [120]. Anterograde signaling governs nuclear genes encoding mitochondrial proteins (NGEMPs) expression to sustain mitochondrial functions and metabolic adaptation, mechanisms co-opted by tumors to meet oncogenic demands [121-124].

Our focus herein centers on mitochondrial retrograde regulation of nuclear activity in oncogenic contexts.

5.1 Retrograde Signaling Pathways

Mitochondrial retrograde signaling (MRS) enables organelle-to-nucleus communication via specific molecular messengers, regulating nuclear gene expression to coordinate stress responses, metabolic adaptation, and environmental acclimatization [125,126].

Ca²⁺ serves as pivotal retrograde mediator - physiological mitochondrial uptake occurs via low-affinity transporters requiring microdomain Ca²⁺ gradients [127], while oncogenic dysfunction disrupts Ca²⁺ homeostasis, triggering pro-survival signaling. Mitochondrial dysfunction leads to increased cytosolic calcium concentration and activates calmodulin, which in turn activates multiple downstream effectors such as nuclear factor-κB (NF-κB), nuclear factor-activated T cells (NFAT), and activated transcription factors (ATF). These transcription factors regulate the expression of genes involved in cell proliferation, survival, and metabolism [125,126]. Pharmacological Hsp90 blockade triggers mitochondrial Ca²⁺ efflux, causing ER stress-mediated CHOP upregulation and tumor cell death [128]. Mitochondrial Ca²⁺ dysregulation activates MAPK/NF-κB/mTOR pathways, enhancing proliferation, invasion, and therapy resistance [129-131]. Oncogenic Ca²⁺ signaling reprograms kinase networks (ERK/IKK/mTORC1) to promote metastatic adaptation and chemoresistance [129-131].

Stress-induced nuclear translocation of mitochondrial proteins mediates direct gene regulation: GPS2 coordinates H3K9 demethylation and Pol II activation [132], while ATFS-1 activates mitochondrial stress response genes [133].

Cytosolic mtDNA activates innate immunity through pattern recognition receptors [134], creating immunogenic microenvironments that influence oncologic outcomes [135,136]. mtDNA liberation constitutes conserved stress response, hyperactivated in malignancies through diverse mechanisms [137]. Tumor microenvironment stressors (metabolic shifts, lipid overload) induce ROS-mediated mtDNA damage and mitophagy, facilitating mtDNA escape via mitochondrial-derived vesicles [138-142]. Pathological IMM permeability enables mtDNA pore-mediated release [137,143], while fumarate accumulation drives MDV-dependent mtDNA export [144]. Tumor-derived EVs transfer mtDNA to immune cells, altering their functional states [142]. Liberated mtDNA activates cGAS-STING signaling [145], reprograms nuclear transcription [135], and triggers damage associated molecular patterns (DAMPs)-mediated immunity [146,147], collectively shaping inflammatory landscapes and oncologic trajectories.

5.2 Metabolite-Mediated Chromatin Remodeling

Mitochondrial metabolites translocate to nuclei as substrates/cofactors for chromatin-modifying enzymes, regulating gene expression through DNA methylation

and histone modifications [148,149]. This metabolism-epigenetics crosstalk provides novel insights into oncogenesis and therapeutic development [150].

Normal cells generate acetyl-CoA via mitochondrial TCA cycle, while cancer cells exhibit profound metabolic rewiring [151,152]. These alterations impact both bioenergetics and histone acetylation patterns through spatial-temporal regulation of acetyl-CoA availability [148,153,154]. Cancer-associated metabolic reprogramming enhances fatty acid oxidation [151,155] and upregulates ATP-citrate synthase (ACLY) [156], optimizing acetyl-CoA production/utilization. Mitochondrial acetyl-CoA converts to acetyl-carnitine via carnitine acetyltransferase (CAT), transported to nuclei and reconverted to acetyl-CoA by nuclear CAT, providing histone acetyltransferase (HAT) substrates for histone acetylation [157-159]. At the same time, cancer cells exhibit dysregulated HAT/histone deacetylase (HDAC) equilibrium, causing pathological histone acetylation patterns [160-162]. Acetylation-induced chromatin decompaction facilitates pioneer factor binding and mediates bromodomain-containing protein recruitment, synergistically activating oncogenic transcription [162,163]. Acetylation-mediated chromatin remodeling regulates oncogenes/tumor suppressors [162,164,165], while HDAC inhibitors modulate cell cycle/apoptosis [160,166,167] and metastatic potential [168] through acetylation dynamics.

S-adenosylmethionine (SAM), synthesized via methionine adenosyltransferase (MAT)-mediated methionine/ATP condensation [169], serves as universal methyl donor for DNA/histone methylation and other biosynthetic processes [170]. Mitochondrial defects in cancer impair one-carbon metabolism [171,172] and ATP production, disrupting SAM biosynthesis. SAM depletion induces genome-wide hypomethylation [173,174], whereas SAM supplementation hypermethylates DNA [175], demonstrating dose-dependent epigenetic regulation. Context-specific SAM depletion enables proto-oncogene activation via promoter demethylation, pharmacologically reversible through methyl donor supplementation [173]. Under SAM deficiency, H3K9 monomethylation preserves heterochromatin integrity and enables epigenetic memory restoration upon metabolic recovery [176]. In hepatocellular carcinoma (HCC), MAT1A downregulation and MAT2A upregulation reduce hepatic SAM levels. MAT1A-KO mice exhibit SAM deficiency with macrovesicular steatosis, monocyte infiltration, and HCC development. Colorectal cancer models show SAM depletion correlates with enhanced proliferation, survival, and angiogenesis [177]. SAM deficiency drives tumor progression through integrated metabolic-enzymatic-epigenetic dysregulation of DNA/histone methylation [178-181]. Tumor methylation dynamics significantly correlate with patient survival outcomes [182].

TCA intermediates α -KG and succinate regulate histone demethylation through competitive modulation of KDMs [183,184]. α -KG is an essential cofactor for a variety of α -KG-dependent dioxygenases [185,186], and α -KG-dependent histone demethylases, such as KDM4A and

JARID1B, are aberrantly expressed in a variety of cancers, affecting gene expression and cellular function [187,188]. Succinate antagonizes α -KG by competitively inhibiting KDMs, stabilizing repressive histone marks [184,189]. Metabolic control of α -KG/succinate balance regulates stem cell fate decisions through TET/KDM-mediated DNA/histone demethylation [190,191]. Pharmacological modulation of α -KG/succinate levels enables precise control of chromatin states and stemness-associated transcription.

Lactate serves as metabolic transmitter coordinating: 1) MDSC polarization, 2) T cell exhaustion, 3) deubiquitinase activation in tumor cells. Histone lactylation - lactoyl-lysine PTM - enhances chromatin accessibility and transcriptional activation [191-193]. Tumor lactate overload drives hyperlactylation, correlating with aggressive phenotypes and poor prognosis across malignancies [194-196]. This epigenetic-metabolic circuit links glycolytic flux to PD-L1 upregulation [197] and MCT4 overexpression [198], enforcing therapeutic resistance.

Mitochondrial calcium dysregulation and mtDNA release drive nuclear transcriptional reprogramming and immune evasion. Mitochondrial-nuclear crosstalk, mediated by metabolite-driven epigenetic remodeling and mtDNA-triggered immunomodulation, serves as a central hub for tumor heterogeneity and immune escape. Targeting this interaction network not only suppresses malignant progression but also reprograms the immune microenvironment, offering a novel paradigm for developing "metabolism-epigenetics-immunity" integrated precision therapies.

6. Mitochondrial Crosstalk with the Tumor Microenvironment

As a heterogeneous milieu, the TME integrates immunocytes, cancer-associated fibroblasts, angiogenic networks, and matrix components [199,200] into self-reinforcing circuits that orchestrate malignant progression through biophysical and biochemical interactions [201]. Mitochondrial-TME interactions operate through metabolic reprogramming, immunomodulation, and paracrine signaling - mechanisms that extend beyond bioenergetic support to govern tumor initiation, dissemination, and therapeutic resistance.

6.1 Mitochondrial Regulation of Immune Evasion and Metastatic Competence

Mitochondria regulate immune cell activation, differentiation, and effector functions through metabolic modulation, ROS production, and cell death regulation [202], thereby shaping immune responses and facilitating tumor immune escape. As mentioned, mitochondrial involvement in energy production confers metabolic flexibility to cancer cells, predominantly relying on glycolysis for bioenergetic demands [202,203]. This specific metabolic mode helps tumor cells to escape the host immune response. On the one hand, the decrease of pH in the tumor microenvironment caused by the

production of large amounts of lactic acid by tumor cells inhibits the function of immune effector cells, affects the metabolism of immune cells, and promotes the production of immunosuppressive cells such as Treg cells [204-206], or the nutrient deficiency in the tumor microenvironment caused by massive glucose consumption affects the activation and effector function of T cells [207]. On the other hand, mitochondrial dysfunction—particularly metabolic reprogramming—directly impacts the expression of immune checkpoint molecules on tumor cells. This occurs through direct genetic regulation and indirect metabolite-mediated epigenetic modulation of PD-1, PD-L1, and CTLA-4 expression [208,209]. Studies demonstrate that epigenetic loss of the ATP synthase subunit ATP5H triggers core metabolic rewiring, leading to ROS accumulation and stabilization of hypoxia-inducible factor 1 α (HIF-1 α) under normoxic conditions [210]. HIF-1 α stabilization subsequently upregulates PD-L1 expression, thereby suppressing T cell-mediated antitumor activity [211]. Horizontal mtDNA transfer from tumor to TILs induces metabolic paralysis and premature senescence in recipient T cells, compromising antitumor immunity through mitochondrial dysfunction propagation [141]. Mitochondrial reprogramming establishes pre-metastatic niches through TME remodeling, priming distant sites for metastatic colonization [212].

6.2 Intercellular Mitochondrial Transfer in the Tumor Microenvironment

Mitochondria can traffic between cells via intercellular transfer mechanisms [213], a phenomenon particularly prevalent in tumor ecosystems that critically influences malignant progression and immune evasion [214]. Neoplastic networks engage in mitochondrial commerce with diverse TME residents - from bone marrow-derived cells to vascular endothelia - through specialized intercellular conduits [215-217]. Acquired mitochondria reprogram recipient cell bioenergetics [218], epigenetic landscapes [219], and survival signaling [220], collectively remodeling TME physicochemical properties to favor malignancy [221-224].

Therapeutic and microenvironmental stresses (ROS, nutrient deprivation, chemotherapeutics) induce mitochondrial dysfunction that triggers compensatory transfer mechanisms in tumors [225,226]. Breast cancer models demonstrate oxidative stress (H₂O₂ exposure) significantly enhances mitochondrial trafficking between malignant and stromal cells [227]. mtDNA damage-induced respiratory collapse (<30% baseline ATP production) triggers mitochondrial vampirism - tumor cells replenish functional mitochondria from adjacent stroma to regain bioenergetic competence [214]. Implanting defective mitochondria (Mito8344/P-Mito8344) into recipient cells induces metabolic reprogramming - glycolytic flux increases 3-fold while oxygen consumption decreases by 60% [227]. There is mitochondrial transfer between stromal cells, immune cells and cancer cells in the tumor microenvironment. Bone marrow stromal cells can transfer mitochondria to myeloma cells by forming tunneling nanotubes (TNTs)

connecting myeloma cells and stromal cells [228], CAFs can allow the exchange of mitochondria and other cargo between cells through contact-dependent tunneling nanotubes (TNTs) with breast cancer cells [229], and nearly unidirectional mitochondria presented by T cells to tumor cells. Somatic metastasis causes "enhanced metabolism" of tumor cells and "depletion" of immune cells [221].

Mitochondrial trafficking employs TNTs [228], EVs [230], and fusogenic mechanisms [215], representing novel therapeutic targets to disrupt metabolic symbiosis and restore treatment sensitivity [231].

7. Prospect: Therapeutic Strategies Targeting Mitochondria

Mitochondria function as an "adaptive engine" in tumor evolution, driving carcinogenesis, metastasis, and therapy resistance through metabolic reprogramming, dynamic regulation, stress signaling, and mitochondrial-nuclear crosstalk. These mechanisms position mitochondrial-targeted therapies as pivotal approaches to overcome limitations of conventional treatments. Mitochondrial-targeted therapeutic strategies aim to selectively eliminate cancer cells by disrupting tumor-specific mitochondrial functions while minimizing off-target effects on normal cells. Current mitochondrial-targeted strategies capitalize on tumor-specific metabolic vulnerabilities, inducing oncogenic metabolic crisis and programmed cell death through selective disruption of energy production, genetic stability, or dynamic equilibrium. Compared to traditional chemotherapy's broad cytotoxicity, these strategies enhance selectivity through dual mechanisms: 1) Cancer cells' heightened mitochondrial activity increases susceptibility to inhibition; 2) Targeted delivery systems exploit tumor mitochondrial properties for spatial drug enrichment, reducing off-target toxicity. Therapeutic objectives extend beyond direct tumor killing to disrupting mitochondrial-mediated chemoresistance mechanisms, thereby reversing drug resistance and enhancing combination therapy efficacy. Current research employs multidimensional interventions: metabolic reprogramming control, bioenergetic machinery disruption, genetic system targeting, and smart delivery technologies to overcome drug accumulation barriers, ultimately establishing precision strike systems against tumor mitochondrial adaptation networks.

Due to the Warburg effect, many cancers exhibit high dependence on glycolysis, making glycolysis targeting a promising anticancer strategy. Recent advances in studying glycolytic enzymes/proteins have identified multiple therapeutic targets: glucose transporters (GLUTs) [232], hexokinase (HK) [233,234], phosphofructokinase (PFK) [235], pyruvate kinase M2 (PKM2) [236], lactate dehydrogenase A (LDHA) [237], and pyruvate dehydrogenase kinase (PDK) [238]. Various natural and synthetic compounds effectively inhibit these glycolytic targets, demonstrating antitumor potential. The electron transport chain (ETC) is central to mitochondrial energetics. Many tumors critically depend on ETC function; its inhibition may selectively kill

cancer cells with minimal normal cell toxicity. Examples include complex I inhibitors metformin and IACS-010759 [239,240], and complex III inhibitor atovaquone [241], which suppress ATP production to induce bioenergetic collapse. Similarly, targeting the TCA cycle represents a viable strategy to inhibit tumor growth and proliferation. CPI-613 targets TCA cycle enzymes pyruvate dehydrogenase complex (PDC) and α -ketoglutarate dehydrogenase (KGDH), reducing metabolic flexibility and depleting nutrient reserves in tumors [238,242]. Pyruvate transport inhibitor UK-5099 disrupts TCA cycle by blocking mitochondrial pyruvate uptake [243]. Modulating TCA intermediates like succinate and their downstream signaling pathways represents another potential therapeutic approach [244]. Enhanced fatty acid synthase (FASN) activity in tumors supports membrane biogenesis and energy storage. FASN inhibition effectively suppresses tumor growth/metastasis [245,246], with clinical candidates including GSK2194069, JNJ-54302833, IPI-9119, and TVB-2640 [247]. FAO provides critical energy for tumors. FAO inhibitors etomoxir and perhexiline demonstrate antitumor efficacy [248,249], while trimetazidine targets mitochondrial trifunctional protein (MTP) [250]. mtDNA-targeting agents induce tumor death via mitochondrial dysfunction, ROS overproduction, and apoptosis. Anthracyclines (doxorubicin) [251], cisplatin [252], topoisomerase I inhibitors (homoharringtonine/topotecan) [253], and Pol γ inhibitors (Congo red) [254] all damage mtDNA.

However, traditional chemotherapeutic agents lack targeting specificity, leading to nonspecific distribution in normal tissues and mitochondrial membrane impermeability, which prevents effective drug accumulation at target sites. To address these limitations, mitochondrial-targeted drug delivery systems have been developed to precisely transport therapeutic agents to cancer cell mitochondria. These systems not only enhance therapeutic efficacy but also reduce off-target toxicity, thereby overcoming the drawbacks of conventional chemotherapy. Leveraging the hyperpolarized mitochondrial membrane potential in cancer cells, positively charged carriers such as triphenylphosphonium (TPP)-modified nanoparticles have been engineered [255,256]. Mitochondria-targeting peptide-based nanoplateforms efficiently deliver anticancer drugs to tumor mitochondria, significantly enhancing antitumor activity [257]. Additionally, liposomes and polymeric nanocarriers serve as critical delivery platforms due to their biocompatibility and tunability [258,259]. Mitochondrial-targeted delivery systems markedly improve chemotherapeutic outcomes [256,260] and demonstrate substantial potential in overcoming multidrug resistance [63,261]. Integration with other modalities like photodynamic therapy (PDT), photothermal therapy (PTT), and chemodynamic therapy further amplifies therapeutic efficacy [262,263].

Current targeting strategies can disrupt tumor energy supply, but metabolic plasticity may activate compensatory pathways. Future therapies should combine multiple metabolic nodes (e.g., mitochondrial-targeted agents + traditional chemotherapeutics) to

synergistically amplify therapeutic stress. Although current delivery systems improve mitochondrial drug accumulation, challenges remain in addressing tumor heterogeneity-induced targeting variability and mitochondrial membrane potential differences affecting drug enrichment. Mitochondrial-targeted therapy will evolve from broad cytotoxicity to precision regulation, integrating metabolism, genetics, immunology, and engineering to establish "diagnosis-treatment-monitoring" precision medicine frameworks. Advances in AI-driven drug design, single-cell metabolomics, and smart delivery systems may enable real-time intervention in mitochondrial networks, offering revolutionary solutions for drug resistance and metastasis. These advancements will transform cancer treatment and potentially provide novel strategies for metabolic disorders and age-related diseases.

Declarations

Ethics Approval and Consent to Participate

Not applicable

Consent for Publication

All authors have approved the manuscript and agreed to submission.

Availability of Data and Material

Not applicable

Competing Interests

The authors declare that they have no competing interests.

Author Contributions

J. Zhang, J. Xu and B. Hu conceived the work, analyzed the data, and drafted the manuscript. All authors contributed to the article and approved the submitted version.

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References

- [1] Walker UA. Mitochondriale Medizin für Internisten [Mitochondrial medicine for internists]. *Medizinische Klinik*. 2000, 95(12), 689-96. DOI: 10.1007/pl00002087
- [2] Frey TG, Mannella CA. The internal structure of mitochondria. *Trends Biochem Sci*. 2000, 25(7), 319-24. DOI: 10.1016/s0968-0004(00)01609-1
- [3] Nunnari J, Suomalainen A. Mitochondria: in sickness and in health. *Cell*. 2012, 148(6), 1145-59. DOI: 10.1016/j.cell.2012.02.035
- [4] Harrington JS, Ryter SW, Plataki M, Price DR, Choi AMK. Mitochondria in health, disease, and aging.

- Physiological Reviews. 2023, 103(4), 2349-2422. DOI: 10.1152/physrev.00058.2021
- [5] Suomalainen A, Nunnari J. Mitochondria at the crossroads of health and disease. *Cell*. 2024, 187(11), 2601-2627. DOI: 10.1016/j.cell.2024.04.037
- [6] Kopinski PK, Singh LN, Zhang SP, Lott MT, Wallace DC. Mitochondrial DNA variation and cancer. *Nature Reviews. Cancer*. 2021, 21(7), 431-445. DOI: 10.1038/s41568-021-00358-w
- [7] Vadakedath S, Kandi V, Ca J, Vijayan S, Achyut KC, Uppuluri S, et al. Mitochondrial Deoxyribonucleic Acid (mtDNA), Maternal Inheritance, and Their Role in the Development of Cancers: A Scoping Review. *Cureus*. 2023, 15(6), e39812. DOI: 10.7759/cureus.39812
- [8] Kozakiewicz P, Grzybowska-Szatkowska L, Ciesielka M, Rzymowska J. The Role of Mitochondria in Carcinogenesis. *International Journal of Molecular Sciences*. 2021, 22(10), 5100. DOI: 10.3390/ijms22105100
- [9] Jafarinia M, Vos T, Lim SS, Naghavi M, Murray CJL, Onwujekwe O. Global burden of 369 diseases and injuries in 204 countries and territories, 1990-2019: a systematic analysis for the Global Burden of Disease Study 2019. *Lancet*. 2020, 396(10258), 1204-1222. DOI: 10.1016/S0140-6736(20)30925-9
- [10] Pegoraro C, Domingo-Ortí I, Conejos-Sánchez I, Vicent MJ. Unlocking the Mitochondria for Nanomedicine-based Treatments: Overcoming Biological Barriers, Improving Designs, and Selecting Verification Techniques. *Advanced Drug Delivery Reviews*. 2024, 207, 115195. DOI: 10.1016/j.addr.2024.115195
- [11] Choudhury FK. Mitochondrial Redox Metabolism: The Epicenter of Metabolism during Cancer Progression. *Antioxidants (Basel)*. 2021, 10(11), 1838. DOI: 10.3390/antiox10111838
- [12] Ciccarone F, Ciriolo MR. Reprogrammed mitochondria: a central hub of cancer cell metabolism. *Biochemical Society Transactions*. 2024, 52(3), 1305-1315. DOI: 10.1042/BST20231090
- [13] Kukurugya MA, Rosset S, Titov DV. The Warburg Effect is the result of faster ATP production by glycolysis than respiration. *Proceedings of the National Academy of Sciences of the United States of America*. 2024, 121(46), e2409509121. DOI: 10.1073/pnas.2409509121
- [14] WARBURG O. On the origin of cancer cells. *Science*. 1956, 123(3191), 309-14. DOI: 10.1126/science.123.3191.309
- [15] Wang T, Sun FZ, Li CC, Nan P, Song Y, Wan XH, et al. MTA1, a Novel ATP Synthase Complex Modulator, Enhances Colon Cancer Liver Metastasis by Driving Mitochondrial Metabolism Reprogramming. *Advanced Science*. 2023, 10(25), e2300756. DOI: 10.1002/advs.202300756
- [16] Lai HW, Kasai M, Yamamoto S, Fukuhara H, Karashima T, Kurabayashi A, et al. Metabolic shift towards oxidative phosphorylation reduces cell-density-induced cancer-stem-cell-like characteristics in prostate cancer in vitro. *Biology Open*. 2023, 12(4), bio059615. DOI: 10.1242/bio.059615
- [17] Wu ZD, Zuo ML, Zeng L, Cui KS, Liu B, Yan CJ, et al. OMA1 reprograms metabolism under hypoxia to promote colorectal cancer development. *EMBO Reports*. 2021, 22(1), e50827. DOI: 10.15252/embr.202050827
- [18] Liu J, Peng YH, Shi L, Wan LX, Inuzuka H, Long JG, et al. Skp2 dictates cell cycle-dependent metabolic oscillation between glycolysis and TCA cycle. *Cell Research*. 2021, 31(1), 80-93. DOI: 10.1038/s41422-020-0372-z
- [19] Zeng SY, Hu X. Lactic acidosis switches cancer cells from dependence on glycolysis to OXPHOS and renders them highly sensitive to OXPHOS inhibitors. *Biochemical and Biophysical Research Communications*. 2023, 671, 46-57. DOI: 10.1016/j.bbrc.2023.05.097
- [20] Smith B, Schafer XL, Ambeskovic A, Spencer CM, Land H, Munger J. Addiction to Coupling of the Warburg Effect with Glutamine Catabolism in Cancer Cells. *Cell Reports*. 2016, 17(3), 821-836. DOI: 10.1016/j.celrep.2016.09.045
- [21] Son J, Lyssiotis CA, Ying HQ, Wang XX, Hua SJ, Ligorio M, et al. Glutamine supports pancreatic cancer growth through a KRAS-regulated metabolic pathway. *Nature*. 2013, 496(7443), 101-5. DOI: 10.1038/nature12040
- [22] Mailloux RJ, Bériault R, Lemire J, Singh R, Chénier DR, Hamel RD, et al. The tricarboxylic acid cycle, an ancient metabolic network with a novel twist. *PLoS One*. 2007, 2(8), e690. DOI: 10.1371/journal.pone.0000690
- [23] Boese AC, Kang S. Mitochondrial metabolism-mediated redox regulation in cancer progression. *Redox Biology*. 2021, 42, 101870. DOI: 10.1016/j.redox.2021.101870
- [24] Vander Heiden MG, Cantley LC, Thompson CB. Understanding the Warburg effect: the metabolic requirements of cell proliferation. *Science*. 2009, 324(5930), 1029-33. DOI: 10.1126/science.1160809
- [25] Filipp FV, Scott DA, Ronai ZA, Osterman AL, Smith JW. Reverse TCA cycle flux through isocitrate dehydrogenases 1 and 2 is required for lipogenesis in hypoxic melanoma cells. *Pigment Cell & Melanoma Research*. 2012, 25(3), 375-83. DOI: 10.1111/j.1755-148X.2012.00989.x
- [26] Faubert B, Vincent EE, Griss T, Samborska B, Izreig S, Svensson RU, et al. Loss of the tumor suppressor LKB1 promotes metabolic reprogramming of cancer cells via HIF-1 α . *Proceedings of the National Academy of Sciences of the United States of America*. 2014, 111(7), 2554-9. DOI: 10.1073/pnas.1312570111
- [27] Guo D, Tong YY, Jiang XM, Meng Y, Jiang HF, Du LY, et al. Aerobic glycolysis promotes tumor immune evasion by hexokinase2-mediated phosphorylation of I κ B α . *Cell Metabolism*. 2022, 34(9), 1312-1324.e6. DOI: 10.1016/j.cmet.2022.08.002
- [28] Fu Y, Zou TT, Shen XT, Nelson PJ, Li JH, Wu C, et al. Lipid metabolism in cancer progression and therapeutic strategies. *MedComm (2020)*. 2020, 2(1), 27-59. DOI: 10.1002/mco2.27
- [29] Riscal R, Skuli N, Simon MC. Even Cancer Cells Watch Their Cholesterol! *Molecular Cell*. 2019, 76(2), 220-231. DOI: 10.1016/j.molcel.2019.09.008
- [30] Cheng CM, Geng F, Cheng X, Guo DL. Lipid metabolism reprogramming and its potential targets in cancer. *Cancer Communications*. 2018, 38(1), 27. DOI: 10.1186/s40880-018-0301-4
- [31] Baenke F, Peck B, Miess H, Schulze A. Hooked on fat: the role of lipid synthesis in cancer metabolism and tumour development. *Disease Models & Mechanisms*. 2013, 6(6), 1353-63. DOI: 10.1242/dmm.011338
- [32] Pellerin L, Carrié L, Dufau C, Nieto L, Séguib B, Levade T, et al. Lipid metabolic Reprogramming: Role in Melanoma Progression and Therapeutic Perspectives. *Cancers (Basel)*. 2020, 12(11), 3147. DOI: 10.3390/cancers12113147
- [33] Parrales A, Iwakuma T. p53 as a Regulator of Lipid Metabolism in Cancer. *International Journal of Molecular Sciences*. 2016, 17(12), 2074. DOI: 10.3390/ijms17122074
- [34] Yang K, Wang XK, Song CH, He Z, Wang RX, Xu YR, et al. The role of lipid metabolic reprogramming in tumor microenvironment. *Theranostics*. 2023, 13(6), 1774-1808. DOI: 10.7150/thno.82920
- [35] Tilokani L, Nagashima S, Paupe V, Prudent J.

- Mitochondrial dynamics: overview of molecular mechanisms. *Essays Biochem.* 2018, 62(3), 341-360. DOI: 10.1042/EBC20170104
- [36] Chen W, Zhao HK, Li YS. Mitochondrial dynamics in health and disease: mechanisms and potential targets. *Signal Transduction and Targeted Therapy.* 2023, 8(1), 333. DOI: 10.1038/s41392-023-01547-9
- [37] Liesa M, Palacín M, Zorzano A. Mitochondrial dynamics in mammalian health and disease. *Physiological Reviews.* 2009, 89(3), 799-845. DOI: 10.1152/physrev.00030.2008
- [38] Zhao J, Zhang J, Yu M, Xie Y, Huang Y, Wolff DW, et al. Mitochondrial dynamics regulates migration and invasion of breast cancer cells. *Oncogene.* 2013, 32(40), 4814-24. DOI: 10.1038/onc.2012.494
- [39] Han SY, Jeong YJ, Choi Y, Hwang SK, Bae YS, Chang YC. Mitochondrial dysfunction induces the invasive phenotype, and cell migration and invasion, through the induction of AKT and AMPK pathways in lung cancer cells. *International Journal of Molecular Medicine.* 2018, 42(3), 1644-1652. DOI:10.3892/ijmm.2018.3733
- [40] Kleele T, Rey T, Winter J, Zaganelli S, Mahecic D, Perreten Lambert H, et al. Distinct fission signatures predict mitochondrial degradation or biogenesis. *Nature.* 2021, 593(7859), 435-439. DOI: 10.1038/s41586-021-03510-6
- [41] Ma JT, Zhang XY, Cao R, Sun L, Jing W, Zhao JZ, et al. Effects of Dynamin-related Protein 1 Regulated Mitochondrial Dynamic Changes on Invasion and Metastasis of Lung Cancer Cells. *Journal of Cancer.* 2019, 10(17), 4045-4053. DOI: 10.7150/jca.29756
- [42] Yang YF, Ouyang YS, Yang LC, Beal MF, McQuibban A, Vogel H, et al. Pink1 regulates mitochondrial dynamics through interaction with the fission/fusion machinery. *Proceedings of the National Academy of Sciences of the United States of America.* 2008, 105(19), 7070-5. DOI: 10.1073/pnas.0711845105
- [43] Li YJ, Chen H, Yang Q, Wan LX, Zhao J, Wu YY, et al. Increased Drp1 promotes autophagy and ESCC progression by mtDNA stress mediated cGAS-STING pathway. *Journal of Experimental & Clinical Cancer Research: CR.* 2022, 41(1), 76. DOI: 10.1186/s13046-022-02262-z
- [44] Yin MJ, Lu Q, Liu X, Wang T, Liu Y, Chen LF. Silencing Drp1 inhibits glioma cells proliferation and invasion by RHOA/ ROCK1 pathway. *Biochemical and Biophysical Research Communications.* 2016, 478(2), 663-8. DOI: 10.1016/j.bbrc.2016.08.003
- [45] Humphries BA, Cutter AC, Buschhaus JM, Chen YC, Qyli T, Palagama DSW, et al. Enhanced mitochondrial fission suppresses signaling and metastasis in triple-negative breast cancer. *Breast Cancer Research: BCR.* 2020, 22(1), 60. DOI: 10.1186/s13058-020-01301-x
- [46] Sánchez-Alvarez R, De Francesco EM, Fiorillo M, Sotgia F, Lisanti MP. Mitochondrial Fission Factor (MFF) Inhibits Mitochondrial Metabolism and Reduces Breast Cancer Stem Cell (CSC) Activity. *Frontiers in Oncology.* 2020, 10, 1776. DOI: 10.3389/fonc.2020.01776
- [47] Parida PK, Marquez-Palencia M, Ghosh S, Khandelwal N, Kim K, Nair V, et al. Limiting mitochondrial plasticity by targeting DRP1 induces metabolic reprogramming and reduces breast cancer brain metastases. *Nature Cancer.* 2023, 893-907. DOI: 10.1038/s43018-023-00563-6
- [48] Wu D, Yang Y, Hou YR, Zhao ZF, Liang N, Yuan P, et al. Increased mitochondrial fission drives the reprogramming of fatty acid metabolism in hepatocellular carcinoma cells through suppression of Sirtuin 1. *Cancer Communications.* 2022, 42(1), 37-55. DOI: 10.1002/cac2.12247
- [49] Ippolito L, Morandi A, Taddei ML, Parri M, Comito G, Iscaro A, et al. Cancer-associated fibroblasts promote prostate cancer malignancy via metabolic rewiring and mitochondrial transfer. *Oncogene.* 2019, 38(27), 5339-5355. DOI:10.1038/s41388-019-0805-7
- [50] Gao S, Hu JJ. Mitochondrial Fusion: The Machineries In and Out. *Trends in Cell Biology.* 2021, 31(1), 62-74. DOI: 10.1016/j.tcb.2020.09.008
- [51] Meeusen SL, Nunnari J. How mitochondria fuse. *Current Opinion in Cell Biology.* 2005, 17(4), 389-94. DOI: 10.1016/j.ceb.2005.06.014
- [52] Griffin EE, Detmer SA, Chan DC. Molecular mechanism of mitochondrial membrane fusion. *Biochimica et Biophysica Acta.* 2006, 1763(5-6), 482-9. DOI: 10.1016/j.bbamcr.2006.02.003
- [53] Mishra P, Carelli V, Manfredi G, Chan DC. Proteolytic cleavage of Opa1 stimulates mitochondrial inner membrane fusion and couples fusion to oxidative phosphorylation. *Cell Metabolism.* 2014, 19(4), 630-41. DOI: 10.1016/j.cmet.2014.03.011
- [54] Archer SL. Mitochondrial dynamics--mitochondrial fission and fusion in human diseases. *The New England Journal of Medicine.* 2013, 369(23), 2236-51. DOI: 10.1056/NEJMr1215233
- [55] Wu ZH, Xiao C, Li F, Huang WB, You FM, Li XK. Mitochondrial fusion-fission dynamics and its involvement in colorectal cancer. *Molecular Oncology.* 2024, 18(5), 1058-1075. DOI: 10.1002/1878-0261.13578
- [56] Wu ZH, Xiao C, Long J, Huang WB, You FM, Li XK. Mitochondrial dynamics and colorectal cancer biology: mechanisms and potential targets. *Cell Communication and Signaling: CCS.* 2024, 22(1), 91. DOI: 10.1186/s12964-024-01490-4
- [57] Yu MF, Nguyen ND, Huang YQ, Lin D, Fujimoto TN, Molkenhine JM, et al. Mitochondrial fusion exploits a therapeutic vulnerability of pancreatic cancer. *JCI Insight.* 2019, 5(16), e126915. DOI: 10.1172/jci.insight.126915
- [58] Zamberlan M, Boeckx A, Muller F, Vinelli F, Ek O, Vianello C, et al. Inhibition of the mitochondrial protein Opa1 curtails breast cancer growth. *Journal of Experimental & Clinical Cancer Research: CR.* 2022, 41(1), 95. DOI: 10.1186/s13046-022-02304-6
- [59] Hill S, Van Remmen H. Mitochondrial stress signaling in longevity: a new role for mitochondrial function in aging. *Redox Biology.* 2014, 2, 936-44. DOI: 10.1016/j.redox.2014.07.005
- [60] Raimundo N. Mitochondrial pathology: stress signals from the energy factory. *Trends in Molecular Medicine.* 2014, 20(5), 282-92. DOI: 10.1016/j.molmed.2014.01.005
- [61] Ralph SJ, Rodríguez-Enríquez S, Neuzil J, Saavedra E, Moreno-Sánchez R. The causes of cancer revisited: "mitochondrial malignancy" and ROS-induced oncogenic transformation - why mitochondria are targets for cancer therapy. *Molecular Aspects of Medicine.* 2010, 31(2), 145-70. DOI: 10.1016/j.mam.2010.02.008
- [62] Balliet RM, Capparelli C, Guido C, Pestell TG, Martinez-Outschoorn UE, Lin Z, et al. Mitochondrial oxidative stress in cancer-associated fibroblasts drives lactate production, promoting breast cancer tumor growth: understanding the aging and cancer connection. *Cell Cycle.* 2011, 10(23), 4065-73. DOI: 10.4161/cc.10.23.18254
- [63] Jin P, Jiang JW, Zhou L, Huang Z, Nice EC, Huang CH, et al. Mitochondrial adaptation in cancer drug resistance: prevalence, mechanisms, and management. *Journal of Hematology & Oncology.* 2022, 15(1), 97. DOI: 10.1186/s13045-022-01313-4
- [64] Sabharwal SS, Schumacker PT. Mitochondrial ROS in cancer: initiators, amplifiers or an Achilles' heel? *Nature Reviews. Cancer.* 2014, 14(11), 709-21. DOI: 10.1038/nrc3803

- [65] Pelicano H, Carney D, Huang P. ROS stress in cancer cells and therapeutic implications. *Drug Resistance Updates: Reviews and Commentaries in Antimicrobial and Anticancer Chemotherapy*. 2004, 7(2), 97-110. DOI: 10.1016/j.drug.2004.01.004
- [66] Wang YW, Qi H, Liu Y, Duan C, Liu XL, Xia T, et al. The double-edged roles of ROS in cancer prevention and therapy. *Theranostics*. 2021, 11(10), 4839-4857. DOI: 10.7150/thno.56747
- [67] Sharma V, Collins LB, Chen TH, Herr N, Takeda S, Sun W, et al. Oxidative stress at low levels can induce clustered DNA lesions leading to NHEJ mediated mutations. *Oncotarget*. 2016, 7(18), 25377-90. DOI: 10.18632/oncotarget.8298
- [68] Pan JS, Hong MZ, Ren JL. Reactive oxygen species: a double-edged sword in oncogenesis. *World Journal of Gastroenterology*. 2009, 15(14), 1702-7. DOI: 10.3748/wjg.15.1702
- [69] Kim BY, Han MJ, Chung AS. Effects of reactive oxygen species on proliferation of Chinese hamster lung fibroblast (V79) cells. *Free Radical Biology & Medicine*. 2001, 30(6), 686-98. DOI: 10.1016/s0891-5849(00)00514-1
- [70] Meloche S, Pouyssegur J. The ERK1/2 mitogen-activated protein kinase pathway as a master regulator of the G1- to S-phase transition. *Oncogene*. 2007, 26(22), 3227-39. DOI: 10.1038/sj.onc.1210414
- [71] Liu SL, Lin X, Shi DY, Cheng J, Wu CQ, Zhang YD. Reactive oxygen species stimulated human hepatoma cell proliferation via cross-talk between PI3-K/PKB and JNK signaling pathways. *Archives of Biochemistry and Biophysics*. 2002, 406(2), 173-82. DOI: 10.1016/s0003-9861(02)00430-7
- [72] Fantozzi A, Gruber DC, Pisarsky L, Heck C, Kunita A, Yilmaz M, et al. VEGF-mediated angiogenesis links EMT-induced cancer stemness to tumor initiation. *Cancer Research*. 2014, 74(5), 1566-75. DOI: 10.1158/0008-5472.CAN-13-1641
- [73] Xia C, Meng Q, Liu LZ, Rojanasakul Y, Wang XR, Jiang BH. Reactive oxygen species regulate angiogenesis and tumor growth through vascular endothelial growth factor. *Cancer Research*. 2007, 67(22), 10823-30. DOI: 10.1158/0008-5472.CAN-07-0783
- [74] Liu LZ, Hu XW, Xia C, He J, Zhou Q, Shi XL, et al. Reactive oxygen species regulate epidermal growth factor-induced vascular endothelial growth factor and hypoxia-inducible factor-1 α expression through activation of AKT and P70S6K1 in human ovarian cancer cells. *Free Radical Biology & Medicine*. 2006, 41(10), 1521-33. DOI: 10.1016/j.freeradbiomed.2006.08.003
- [75] Ushio-Fukai M. Redox signaling in angiogenesis: role of NADPH oxidase. *Cardiovascular Research*. 2006, 71(2), 226-35. DOI: 10.1016/j.cardiores.2006.04.015
- [76] Dandapat A, Hu C, Sun L, Mehta JL. Small concentrations of oxLDL induce capillary tube formation from endothelial cells via LOX-1-dependent redox-sensitive pathway. *Arteriosclerosis, Thrombosis, and Vascular Biology*. 2007, 27(11), 2435-42. DOI: 10.1161/ATVBAHA.107.152272
- [77] Lee SY, Jeong EK, Ju MK, Jeon HM, Kim MY, Kim CH, et al. Induction of metastasis, cancer stem cell phenotype, and oncogenic metabolism in cancer cells by ionizing radiation. *Molecular Cancer*. 2017, 16(1), 10. DOI: 10.1186/s12943-016-0577-4
- [78] Huang R, Chen H, Liang JY, Li Y, Yang JL, Luo C, et al. Dual Role of Reactive Oxygen Species and their Application in Cancer Therapy. *Journal of Cancer*. 2021, 12(18), 5543-5561. DOI: 10.7150/jca.54699
- [79] Kennel KB, Gretten FR. Immune cell - produced ROS and their impact on tumor growth and metastasis. *Redox Biology*. 2021, 42, 101891. DOI: 10.1016/j.redox.2021.101891
- [80] Shah R, Ibis B, Kashyap M, Boussiotis VA. The role of ROS in tumor infiltrating immune cells and cancer immunotherapy. *Metabolism: Clinical and Experimental*. 2024, 151, 155747. DOI: 10.1016/j.metabol.2023.155747. Epub 2023 Nov 30
- [81] Xiang HD, Ramil CP, Hai J, Zhang CS, Wang HJ, Watkins AA, et al. Cancer-Associated Fibroblasts Promote Immunosuppression by Inducing ROS-Generating Monocytic MDSCs in Lung Squamous Cell Carcinoma. *Cancer Immunology Research*. 2020, 8(4), 436-450. DOI: 10.1158/2326-6066.CIR-19-0507
- [82] Shimura T, Sasatani M, Kawai H, Kamiya K, Kobayashi J, Komatsu K, Kunugita N. Radiation-Induced Myofibroblasts Promote Tumor Growth via Mitochondrial ROS-Activated TGF β Signaling. *Molecular Cancer Research: MCR*. 2018, 16(11), 1676-1686. DOI: 10.1158/1541-7786.MCR-18-0321
- [83] Kurata S. Selective activation of p38 MAPK cascade and mitotic arrest caused by low level oxidative stress. *The Journal of Biological Chemistry*. 2000, 275(31), 23413-6. DOI: 10.1074/jbc.C000308200
- [84] Lee SO, Joo SH, Kwak AW, Lee MH, Seo JH, Cho SS, et al. Podophyllotoxin Induces ROS-Mediated Apoptosis and Cell Cycle Arrest in Human Colorectal Cancer Cells via p38 MAPK Signaling. *Biomolecules & Therapeutics*. 2021, 29(6), 658-666. DOI: 10.4062/biomolther.2021.143
- [85] Kwak AW, Lee MJ, Lee MH, Yoon G, Cho SS, Chae JL, et al. The 3-deoxysappanchalcone induces ROS-mediated apoptosis and cell cycle arrest via JNK/p38 MAPKs signaling pathway in human esophageal cancer cells. *Phytomedicine: International Journal of Phytotherapy and Phytopharmacology*. 2021, 86, 153564. DOI: 10.1016/j.phymed.2021.153564
- [86] Orrenius S, Gogvadze V, Zhivotovsky B. Mitochondrial oxidative stress: implications for cell death. *Annual Review of Pharmacology and Toxicology*. 2007, 47, 143-83. DOI: 10.1146/annurev.pharmtox.47.120505.105122
- [87] Kagan VE, Tyurin VA, Jiang J, Tyurina YY, Ritov VB, Amoscato AA, et al. Cytochrome c acts as a cardiolipin oxygenase required for release of proapoptotic factors. *Nature Chemical Biology*. 2005, 1(4), 223-32. DOI: 10.1038/nchembio727
- [88] Wu HY, Huang CH, Lin YH, Wang CC, Jan TR. Cannabidiol induced apoptosis in human monocytes through mitochondrial permeability transition pore-mediated ROS production. *Free Radical Biology & Medicine*. 2018, 124, 311-318. DOI: 10.1016/j.freeradbiomed.2018.06.023
- [89] Madesh M, Hajnoczky G. VDAC-dependent permeabilization of the outer mitochondrial membrane by superoxide induces rapid and massive cytochrome c release. *The Journal of Cell Biology*. 2001, 155(6), 1003-15. DOI: 10.1083/jcb.200105057
- [90] Chen ZX, Pervaiz S. Bcl-2 induces pro-oxidant state by engaging mitochondrial respiration in tumor cells. *Cell Death and Differentiation*. 2007, 14(9), 1617-27. DOI: 10.1038/sj.cdd.4402165
- [91] Chen ZX, Pervaiz S. Involvement of cytochrome c oxidase subunits Va and Vb in the regulation of cancer cell metabolism by Bcl-2. *Cell Death and Differentiation*. 2010, 17(3), 408-20. DOI: 10.1038/cdd.2009.132
- [92] Kang YH, Lee SJ. The role of p38 MAPK and JNK in Arsenic trioxide-induced mitochondrial cell death in human cervical cancer cells. *Journal of Cellular Physiology*. 2008, 217(1), 23-33. DOI: 10.1002/jcp.21470
- [93] Tsang WP, Chau SP, Kong SK, Fung KP, Kwok TT. Reactive oxygen species mediate doxorubicin induced p53-independent apoptosis. *Life Sciences*. 2003, 73(16),

- 2047-58. DOI: 10.1016/s0024-3205(03)00566-6
- [94] Ma YF, Zhang J, Zhang Q, Chen P, Song JY, Yu SJ, et al. Adenosine induces apoptosis in human liver cancer cells through ROS production and mitochondrial dysfunction. *Biochemical and Biophysical Research Communications*. 2014, 448(1), 8-14. DOI: 10.1016/j.bbrc.2014.04.007
- [95] Liu Y, Borchert GL, Surazynski A, Hu CA, Phang JM. Proline oxidase activates both intrinsic and extrinsic pathways for apoptosis: the role of ROS/superoxides, NFAT and MEK/ERK signaling. *Oncogene*. 2006, 25(41), 5640-7. DOI: 10.1038/sj.onc.1209564
- [96] Chaudhary P, Vishwanatha JK. c-Jun NH2-terminal kinase-induced proteasomal degradation of c-FLIPL/S and Bcl2 sensitize prostate cancer cells to Fas- and mitochondria-mediated apoptosis by tetrandrine. *Biochemical Pharmacology*. 2014, 91(4), 457-73. DOI: 10.1016/j.bcp.2014.08.014
- [97] Gogna R, Madan E, Keppler B, Pati U. Gallium compound GaQ(3) -induced Ca(2+) signalling triggers p53-dependent and -independent apoptosis in cancer cells. *British Journal of Pharmacology*. 2012, 166(2), 617-36. DOI: 10.1111/j.1476-5381.2011.01780.x
- [98] Fujii S, Ushioda R, Nagata K. Redox states in the endoplasmic reticulum directly regulate the activity of calcium channel, inositol 1,4,5-trisphosphate receptors. *Proceedings of the National Academy of Sciences of the United States of America*. 2023, 120(22), e2216857120. DOI: 10.1073/pnas.2216857120
- [99] Hamilton S, Terentyeva R, Martin B, Perger F, Li J, Stepanov A, et al. Increased RyR2 activity is exacerbated by calcium leak-induced mitochondrial ROS. *Basic Research in Cardiology*. 2020, 115(4), 38. DOI: 10.1007/s00395-020-0797-z
- [100] Zeng YQ, Du QD, Zhang ZW, Ma J, Han L, Wang YY, et al. Curcumin promotes cancer-associated fibroblasts apoptosis via ROS-mediated endoplasmic reticulum stress. *Archives of Biochemistry and Biophysics*. 2020, 694, 108613. DOI: 10.1016/j.abb.2020.108613
- [101] Lin YN, Jiang M, Chen WJ, Zhao TJ, Wei YF. Cancer and ER stress: Mutual crosstalk between autophagy, oxidative stress and inflammatory response. *Biomed Pharmacother*. 2019, 118, 109249. DOI: 10.1016/j.biopha.2019.109249
- [102] Li LW, Xing ZH, Wang JY, Guo YH, Wu XM, Ma YM, et al. Hyaluronic acid-mediated targeted nano-modulators for activation of pyroptosis for cancer therapy through multichannel regulation of Ca²⁺ overload. *International Journal of Biological Macromolecules*. 2025, 299, 140116. DOI: 10.1016/j.ijbiomac.2025.140116
- [103] Yang XH, Jiang JT, Yang XY, Han JC, Zheng QS. Licochalcone A induces T24 bladder cancer cell apoptosis by increasing intracellular calcium levels. *Molecular Medicine Reports*. 2016, 14(1), 911-919. DOI: 10.3892/mmr.2016.5334
- [104] Nakagawa T, Zhu H, Morishima N, Li E, Xu J, Yankner BA, et al. Caspase-12 mediates endoplasmic-reticulum-specific apoptosis and cytotoxicity by amyloid-beta. *Nature*. 2000, 403(6765), 98-103. DOI: 10.1038/47513
- [105] Yang J, Zhao XY, Tang M, Li L, Lei Y, Cheng P, et al. The role of ROS and subsequent DNA-damage response in PUMA-induced apoptosis of ovarian cancer cells. *Oncotarget*. 2017, 8(14), 23492-23506. DOI: 10.18632/oncotarget.15626
- [106] Zhou ZX, Fan YM, Zong RK, Tan K. The mitochondrial unfolded protein response: A multitasking giant in the fight against human diseases. *Ageing Research Reviews*. 2022, 81, 101702. DOI: 10.1016/j.arr.2022.101702
- [107] Zhu L, Zhou QL, He L, Chen LX. Mitochondrial unfolded protein response: An emerging pathway in human diseases. *Free Radical Biology & Medicine*. 2021, 163, 125-134. DOI: 10.1016/j.freeradbiomed.2020.12.013
- [108] Torres AK, Fleischhart V, Inestrosa NC. Mitochondrial unfolded protein response (UPRmt): what we know thus far. *Frontiers in Cell and Developmental Biology*. 2024, 12, 1405393. DOI: 10.3389/fcell.2024.1405393
- [109] Naresh NU, Haynes CM. Signaling and Regulation of the Mitochondrial Unfolded Protein Response. *Cold Spring Harbor Perspectives in Biology*. 2019, 11(6), a033944. DOI: 10.1101/cshperspect.a033944
- [110] Inigo JR, Chandra D. The mitochondrial unfolded protein response (UPRmt): shielding against toxicity to mitochondria in cancer. *Journal of Hematology & Oncology*. 2022, 15(1), 98. DOI: 10.1186/s13045-022-01317-0
- [111] Zhang XY, Fan YM, Tan K. A bird's eye view of mitochondrial unfolded protein response in cancer: mechanisms, progression and further applications. *Cell Death & Disease*. 2024, 15(9), 667. DOI: 10.1038/s41419-024-07049-y
- [112] Kenny TC, Craig AJ, Villanueva A, Germain D. Mitohormesis Primes Tumor Invasion and Metastasis. *Cell Reports*. 2019, 27(8), 2292-2303.e6. DOI: 10.1016/j.celrep.2019.04.095
- [113] An H, Zhou B, Hayakawa K, Durán Laforet V, Park JH, Nakamura Y, et al. ATF5-Mediated Mitochondrial Unfolded Protein Response (UPRmt) Protects Neurons Against Oxygen-Glucose Deprivation and Cerebral Ischemia. *Stroke*. 2024, 55(7), 1904-1913. DOI: 10.1161/STROKEAHA.123.045550
- [114] Sutandy FXR, Göbner I, Tascher G, Münch C. A cytosolic surveillance mechanism activates the mitochondrial UPR. *Nature*. 2023, 618(7966), 849-854. DOI: 10.1038/s41586-023-06142-0
- [115] Kumar R, Chaudhary AK, Woytash J, Inigo JR, Gokhale AA, Bshara W, et al. A mitochondrial unfolded protein response inhibitor suppresses prostate cancer growth in mice via HSP60. *The Journal of Clinical Investigation*. 2022, 132(13), e149906. DOI: 10.1172/JCI149906
- [116] Kenny TC, Gomez ML, Germain D. Mitohormesis, UPRmt, and the Complexity of Mitochondrial DNA Landscapes in Cancer. *Cancer Research*. 2019, 79(24), 6057-6066. DOI: 10.1158/0008-5472.CAN-19-1395
- [117] Schmidt O, Pfanner N, Meisinger C. Mitochondrial protein import: from proteomics to functional mechanisms. *Nature reviews. Molecular Cell Biology*. 2010, 11(9), 655-657. DOI: 10.1038/nrm2959
- [118] Cannino G, Di Liegro CM, Rinaldi AM. Nuclear-mitochondrial interaction. *Mitochondrion*. 2007, 7(6), 359-366. DOI: 10.1016/j.mito.2007.07.001
- [119] Quirós PM, Mottis A, Auwerx J. Mitonuclear communication in homeostasis and stress. *Nature Reviews. Molecular Cell Biology*. 2016, 17(4), 213-226. DOI: 10.1038/nrm.2016.23
- [120] Mottis A, Herzig S, Auwerx J. Mitocellular communication: Shaping health and disease. *Science*. 2019, 366(6467), 827-832. DOI: 10.1126/science.aax3768
- [121] Ng S, De Clercq I, Van Aken O, Law SR, Ivanova A, Willems P, et al. Anterograde and retrograde regulation of nuclear genes encoding mitochondrial proteins during growth, development, and stress. *Molecular Plant*. 2014, 7(7), 1075-1093. DOI: 10.1093/mp/ssu037
- [122] Popov LD. Mitochondria as intracellular signalling organelles. An update. *Cellular Signalling*. 2023, 109, 110794. DOI: 10.1016/j.cellsig.2023.110794
- [123] Rinaldi L, Delle Donne R, Borzacchiello D, Insabato L, Feliciello A. The role of compartmentalized signaling pathways in the control of mitochondrial activities in cancer cells. *Biochimica et Biophysica Acta. Reviews on*

- Cancer. 2018, 1869(2), 293-302. DOI: 10.1016/j.bbcan.2018.04.004
- [124] Shteinifer-Kuzmine A, Verma A, Arif T, Aizenberg O, Paul A, Shoshan-Barmaz V. Mitochondria and nucleus cross-talk: Signaling in metabolism, apoptosis, and differentiation, and function in cancer. *IUBMB Life*. 2021, 73(3), 492-510. DOI: 10.1002/iub.2407
- [125] Butow RA, Avadhani NG. Mitochondrial signaling: the retrograde response. *Molecular Cell*. 2004, 14(1), 1-15. DOI: 10.1016/s1097-2765(04)00179-0
- [126] Guha M, Avadhani NG. Mitochondrial retrograde signaling at the crossroads of tumor bioenergetics, genetics and epigenetics. *Mitochondrion*. 2013, 13(6), 577-591. DOI: 10.1016/j.mito.2013.08.007
- [127] Brini M. Ca²⁺ signalling in mitochondria: mechanism and role in physiology and pathology. *Cell Calcium*. 2003, 34(4-5), 399-405. DOI: 10.1016/s0143-4160(03)00145-3
- [128] Park HK, Lee JE, Lim J, Kang BH. Mitochondrial Hsp90s suppress calcium-mediated stress signals propagating from mitochondria to the ER in cancer cells. *Molecular Cancer*. 2014, 13, 148. DOI: 10.1186/1476-4598-13-148
- [129] Amuthan G, Biswas G, Ananatheerthavarada HK, Vijayarathay C, Shephard HM, Avadhani NG. Mitochondrial stress-induced calcium signaling, phenotypic changes and invasive behavior in human lung carcinoma A549 cells. *Oncogene*. 2002, 21(51), 7839-7849. DOI: 10.1038/sj.onc.1205983
- [130] Lee YK, Yi EY, Park SY, Jang WJ, Han YS, Jegal ME, et al. Mitochondrial dysfunction suppresses p53 expression via calcium-mediated nuclear factor- κ B signaling in HCT116 human colorectal carcinoma cells. *BMB Reports*. 2018, 51(6), 296-301. DOI: 10.5483/bmbrep.2018.51.6.232
- [131] Rodríguez-Hernández MA, de la Cruz-Ojeda P, López-Grueso MJ, Navarro-Villarán E, Requejo-Aguilar R, Castejón-Vega B, et al. Integrated molecular signaling involving mitochondrial dysfunction and alteration of cell metabolism induced by tyrosine kinase inhibitors in cancer. *Redox Biology*. 2020, 36, 101510. DOI: 10.1016/j.redox.2020.101510
- [132] Cardamone MD, Tanasa B, Cederquist CT, Huang J, Mahdavian K, Li W, et al. Mitochondrial Retrograde Signaling in Mammals Is Mediated by the Transcriptional Cofactor GPS2 via Direct Mitochondria-to-Nucleus Translocation. *Molecular Cell*. 2018, 69(5), 757-772.e7. DOI: 10.1016/j.molcel.2018.01.037
- [133] Nargund AM, Pellegrino MW, Fiorese CJ, Baker BM, Haynes CM. Mitochondrial import efficiency of ATFS-1 regulates mitochondrial UPR activation. *Science*. 2012, 337(6094), 587-590. DOI: 10.1126/science.1223560
- [134] Heilig R, Lee J, Tait SWG. Mitochondrial DNA in cell death and inflammation. *Biochemical Society Transactions*. 2023, 51(1), 457-472. DOI: 10.1042/BST20221525
- [135] Xia LT, Yan XL, Zhang H. Mitochondrial DNA-activated cGAS-STING pathway in cancer: Mechanisms and therapeutic implications. *Biochimica et Biophysica Acta. Reviews on Cancer*. 2025, 1880(1), 189249. DOI: 10.1016/j.bbcan.2024.189249
- [136] Wang SF, Tseng LM, Lee HC. Role of mitochondrial alterations in human cancer progression and cancer immunity. *Journal of Biomedical Science*. 202, 30(1), 61. DOI: 10.1186/s12929-023-00956-w
- [137] Newman LE, Shadel GS. Mitochondrial DNA Release in Innate Immune Signaling. *Annual Review of Biochemistry*. 2023, 92, 299-332. DOI: 10.1146/annurev-biochem-032620-104401
- [138] Singh RK, Srivastava A, Kalaiarasan P, Manvati S, Chopra R, Bamezai RN. mtDNA germ line variation mediated ROS generates retrograde signaling and induces pro-cancerous metabolic features. *Scientific Reports*. 2014, 4, 6571. DOI: 10.1038/srep06571
- [139] Koenig A, Buskiewicz-Koenig IA. Redox Activation of Mitochondrial DAMPs and the Metabolic Consequences for Development of Autoimmunity. *Antioxidants & Redox Signaling*. 2022, 36(7-9), 441-461. DOI: 10.1089/ars.2021.0073
- [140] Li WT, Li YT, Zhao J, Liao JB, Wen WB, Chen Y, et al. Release of damaged mitochondrial DNA: A novel factor in stimulating inflammatory response. *Pathology, Research and Practice*. 2024, 258, 155330. DOI: 10.1016/j.prp.2024.155330
- [141] Ikeda H, Kawase K, Nishi T, Watanabe T, Takenaga K, Inozume T, et al. Immune evasion through mitochondrial transfer in the tumour microenvironment. *Nature*. 2025, 639(8053), E5. DOI: 10.1038/s41586-024-08439-0
- [142] Zhou Z, Qu CH, Zhou PJ, Zhou Q, Li D, Wu X, et al. Extracellular vesicles activated cancer-associated fibroblasts promote lung cancer metastasis through mitophagy and mtDNA transfer. *Journal of Experimental & Clinical Cancer Research: CR*. 2024, 43(1), 158. DOI: 10.1186/s13046-024-03077-w
- [143] Patrushev M, Kasymov V, Patrusheva V, Ushakova T, Gogvadze V, Gaziev A. Mitochondrial permeability transition triggers the release of mtDNA fragments. *Cellular and Molecular Life Sciences: CMLS*. 2004, 61(24), 3100-3103. DOI: 10.1007/s00018-004-4424-1
- [144] Zecchini V, Paupe V, Herranz-Montoya I, Janssen J, Wortel IMN, Morris JL, et al. Fumarate induces vesicular release of mtDNA to drive innate immunity. *Nature*. 2023, 615(7952), 499-506. DOI: 10.1038/s41586-023-05770-w
- [145] Jibril A, Hellmich C, Wojtowicz EE, Hampton K, Maynard R, De Silva R, et al. Plasma cell-derived mtDAMPs activate the macrophage STING pathway, promoting myeloma progression. *Blood*. 2023, 141(25), 3065-3077. DOI: 10.1182/blood.2022018711
- [146] Guha M, Srinivasan S, Ruthel G, Kashina AK, Carstens RP, Mendoza A, et al. Mitochondrial retrograde signaling induces epithelial-mesenchymal transition and generates breast cancer stem cells. *Oncogene*. 2014, 33(45), 5238-5250. DOI: 10.1038/onc.2013.467
- [147] Liu S, Feng M, Guan WX. Mitochondrial DNA sensing by STING signaling participates in inflammation, cancer and beyond. *International Journal of Cancer*. 2016, 139(4), 736-741. DOI: 10.1002/ijc.30074
- [148] Liu X, Chen C, Wang XY, Sun YH, Zhang J, Chen JX, et al. An Epigenetic Role of Mitochondria in Cancer. *Cells*. 2022, 11(16), 2518. DOI: 10.3390/cells11162518
- [149] Santos JH. Mitochondria signaling to the epigenome: A novel role for an old organelle. *Free Radical Biology & Medicine*. 2021, 170, 59-69. DOI: 10.1016/j.freeradbiomed.2020.11.016
- [150] Campbell SL, Wellen KE. Metabolic Signaling to the Nucleus in Cancer. *Molecular Cell*. 2018, 71(3), 398-408. DOI: 10.1016/j.molcel.2018.07.015
- [151] Corbet C, Pinto A, Martherus R, Santiago de Jesus JP, Polet F, Feron O. Acidosis Drives the Reprogramming of Fatty Acid Metabolism in Cancer Cells through Changes in Mitochondrial and Histone Acetylation. *Cell Metabolism*. 2016, 24(2), 311-323. DOI: 10.1016/j.cmet.2016.07.003
- [152] Ohshima K, Oi R, Nojima S, Morii E. Mitochondria govern histone acetylation in colorectal cancer. *The Journal of Pathology*. 2022, 256(2), 164-173. DOI: 10.1002/path.5818
- [153] Lee JV, Carrer A, Shah S, Snyder NW, Wei S, Venneti S, et al. Akt-dependent metabolic reprogramming regulates tumor cell histone acetylation. *Cell Metabolism*. 2014, 20(2), 306-319. DOI: 10.1016/j.cmet.2014.06.004

- [154] He WJ, Li QG, Li XX. Acetyl-CoA regulates lipid metabolism and histone acetylation modification in cancer. *Biochimica et Biophysica Acta. Reviews on Cancer*. 2023, 1878(1), 188837. DOI: 10.1016/j.bbcan.2022.188837
- [155] Han LM, Zhang CY, Wang DN, Zhang JQ, Tang QQ, Li MJ, et al. Retrograde regulation of mitochondrial fission and epithelial to mesenchymal transition in hepatocellular carcinoma by GCN5L1. *Oncogene*. 2023, 42(13), 1024-1037. DOI: 10.1038/s41388-023-02621-w
- [156] Hu XW, Wang DD, Sun LY, Gao Y, Zhou DZ, Tong XM, et al. Disturbed mitochondrial acetylation in accordance with the availability of acetyl groups in hepatocellular carcinoma. *Mitochondrion*. 2021, 60, 150-159. DOI: 10.1016/j.mito.2021.08.004
- [157] Madiraju P, Pande SV, Prentki M, Madiraju SR. Mitochondrial acetylcarnitine provides acetyl groups for nuclear histone acetylation. *Epigenetics*. 2009, 4(6), 399-403. DOI: 10.4161/epi.4.6.9767
- [158] Giangregorio N, Tonazzi A, Console L, Indiveri C. Post-translational modification by acetylation regulates the mitochondrial carnitine/acylcarnitine transport protein. *Molecular and Cellular Biochemistry*. 2017, 426(1-2), 65-73. DOI: 10.1007/s11010-016-2881-0
- [159] Takahashi H, McCaffery JM, Irizarry RA, Boeke JD. Nucleocytosolic acetyl-coenzyme A synthetase is required for histone acetylation and global transcription. *Molecular Cell*. 2006, 23(2), 207-217. DOI: 10.1016/j.molcel.2006.05.040
- [160] Chrun ES, Modolo F, Daniel FI. Histone modifications: A review about the presence of this epigenetic phenomenon in carcinogenesis. *Pathology, Research and Practice*. 2017, 213(11), 1329-1339. DOI: 10.1016/j.prp.2017.06.013
- [161] Fraga MF, Ballestar E, Villar-Garea A, Boix-Chornet M, Espada J, Schotta G, et al. Loss of acetylation at Lys16 and trimethylation at Lys20 of histone H4 is a common hallmark of human cancer. *Nature Genetics*. 2005, 37(4), 391-400. DOI: 10.1038/ng1531
- [162] Neganova ME, Klochkov SG, Aleksandrova YR, Aliev G. Histone modifications in epigenetic regulation of cancer: Perspectives and achieved progress. *Seminars in Cancer Biology*. 2022, 83, 452-471. DOI: 10.1016/j.semcancer.2020.07.015
- [163] Zhao D, Li FL, Cheng ZL, Lei QY. Impact of acetylation on tumor metabolism. *Molecular & Cellular Oncology*. 2014, 1(3), e963452. DOI: 10.4161/23723548.2014.963452
- [164] Miziak P, Baran M, Borkiewicz L, Trombik T, Stepulak A. Acetylation of Histone H3 in Cancer Progression and Prognosis. *International Journal of Molecular Sciences*. 2024, 25(20), 10982. DOI: 10.3390/ijms252010982
- [165] Ono S, Oue N, Kuniyasu H, Suzuki T, Ito R, Matsusaki K, et al. Acetylated histone H4 is reduced in human gastric adenomas and carcinomas. *Journal of Experimental & Clinical Cancer Research: CR*. 2002, 21(3), 377-382. PMID: 12385581.
- [166] Kanao K, Mikami S, Mizuno R, Shinojima T, Murai M, Oya M. Decreased acetylation of histone H3 in renal cell carcinoma: a potential target of histone deacetylase inhibitors. *The Journal of Urology*. 2008, 180(3), 1131-6. DOI: 10.1016/j.juro.2008.04.136
- [167] Leggatt GR, Gabrielli B. Histone deacetylase inhibitors in the generation of the anti-tumour immune response. *Immunology and Cell Biology*. 2012, 90(1), 33-38. DOI: 10.1038/icb.2011.94
- [168] Yasui W, Oue N, Ono S, Mitani Y, Ito R, Nakayama H. Histone acetylation and gastrointestinal carcinogenesis. *Annals of the New York Academy of Sciences*. 2003, 983, 220-231. DOI: 10.1111/j.1749-6632.2003.tb05977.x
- [169] Mato JM, Alvarez L, Ortiz P, Pajares MA. S-adenosylmethionine synthesis: molecular mechanisms and clinical implications. *Pharmacology & Therapeutics*. 1997, 73(3), 265-280. DOI: 10.1016/s0163-7258(96)00197-0
- [170] Ouyang Y, Wu Q, Li JJ, Sun S, Sun SR. S-adenosylmethionine: A metabolite critical to the regulation of autophagy. *Cell Proliferation*. 2020, 53(11), e12891. DOI: 10.1111/cpr.12891
- [171] Rosenberger FA, Moore D, Atanassov I, Moedas MF, Clemente P, Végvári Á, et al. The one-carbon pool controls mitochondrial energy metabolism via complex I and iron-sulfur clusters. *Science Advances*. 2021, 7(8), eabf0717. DOI: 10.1126/sciadv.abf0717
- [172] Lozoya OA, Martinez-Reyes I, Wang TY, Grenet D, Bushel P, Li JY, et al. Mitochondrial nicotinamide adenine dinucleotide reduced (NADH) oxidation links the tricarboxylic acid (TCA) cycle with methionine metabolism and nuclear DNA methylation. *PLoS Biology*. 2018, 16(4), e2005707. DOI: 10.1371/journal.pbio.2005707
- [173] Luo J, Li YN, Wang F, Zhang WM, Geng X. S-adenosylmethionine inhibits the growth of cancer cells by reversing the hypomethylation status of c-myc and H-ras in human gastric cancer and colon cancer. *International Journal of Biological Sciences*. 2010, 6(7), 784-795. DOI: 10.7150/ijbs.6.784
- [174] Nohara K, Baba T, Murai H, Kobayashi Y, Suzuki T, Tateishi Y, et al. Global DNA methylation in the mouse liver is affected by methyl deficiency and arsenic in a sex-dependent manner. *Archives of Toxicology*. 2011, 85(6), 653-661. DOI: 10.1007/s00204-010-0611-z
- [175] Schmidt T, Leha A, Salinas-Riester G. Treatment of prostate cancer cells with S-adenosylmethionine leads to genome-wide alterations in transcription profiles. *Gene*. 2016, 595(2), 161-167. DOI: 10.1016/j.gene.2016.09.032
- [176] Haws SA, Yu DY, Ye CQ, Wille CK, Nguyen LC, Krautkramer KA, et al. Methyl-Metabolite Depletion Elicits Adaptive Responses to Support Heterochromatin Stability and Epigenetic Persistence. *Molecular Cell*. 2020, 78(2), 210-223.e8. DOI: 10.1016/j.molcel.2020.03.004
- [177] Frau M, Feo F, Pascale RM. Pleiotropic effects of methionine adenosyltransferases deregulation as determinants of liver cancer progression and prognosis. *Journal of Hepatology*. 2013, 59(4), 830-841. DOI: 10.1016/j.jhep.2013.04.031
- [178] Mentch SJ, Mehrmohamadi M, Huang L, Liu X, Gupta D, Mattocks D, et al. Histone Methylation Dynamics and Gene Regulation Occur through the Sensing of One-Carbon Metabolism. *Cell Metabolism*. 2015, 22(5), 861-873. DOI: 10.1016/j.cmet.2015.08.024
- [179] Shyh-Chang N, Locasale JW, Lyssiotis CA, Zheng YX, Teo RY, Ratanasirintrao S, et al. Influence of threonine metabolism on S-adenosylmethionine and histone methylation. *Science*. 2013, 339(6116), 222-226. DOI: 10.1126/science.1226603
- [180] Ye CQ, Sutter BM, Wang Y, Kuang Z, Tu BP. A Metabolic Function for Phospholipid and Histone Methylation. *Molecular Cell*. 2017, 66(2), 180-193.e8. DOI: 10.1016/j.molcel.2017.02.026
- [181] Mentch SJ, Locasale JW. One-carbon metabolism and epigenetics: understanding the specificity. *Annals of the New York Academy of Sciences*. 2016, 1363(1), 91-98. DOI: 10.1111/nyas.12956
- [182] Idaghdour Y, Hodgkinson A. Integrated genomic analysis of mitochondrial RNA processing in human cancers. *Genome Medicine*. 2017, 9(1), 36. DOI: 10.1186/s13073-017-0426-0
- [183] Salminen A, Kauppinen A, Hiltunen M, Kaarniranta K. Krebs cycle intermediates regulate DNA and histone

- methylation: epigenetic impact on the aging process. *Ageing Research Reviews*. 2014, 16, 45-65. DOI: 10.1016/j.arr.2014.05.004
- [184] Xiao MT, Yang H, Xu W, Ma SH, Lin HP, Zhu HG, et al. Inhibition of α -KG-dependent histone and DNA demethylases by fumarate and succinate that are accumulated in mutations of FH and SDH tumor suppressors. *Genes & Development*. 2012, 26(12), 1326-1338. DOI: 10.1101/gad.191056.112
- [185] Tsukada Y, Fang J, Erdjument-Bromage H, Warren ME, Borchers CH, Tempst P, et al. Histone demethylation by a family of JmjC domain-containing proteins. *Nature*. 2006, 439(7078), 811-816. DOI: 10.1038/nature04433
- [186] Soloveyichik M, Xu MS, Zaslaver O, Lee K, Narula A, Jiang R, et al. Mitochondrial control through nutritionally regulated global histone H3 lysine-4 demethylation. *Scientific Reports*. 2016, 6, 37942. DOI: 10.1038/srep37942
- [187] Xiang Y, Zhu ZQ, Han G, Ye XL, Xu B, Peng ZC, et al. JARID1B is a histone H3 lysine 4 demethylase up-regulated in prostate cancer. *Proceedings of the National Academy of Sciences of the United States of America*. 2007, 104(49), 19226-19231. DOI: 10.1073/pnas.0700735104
- [188] Young NL, Dere R. Mechanistic insights into KDM4A driven genomic instability. *Biochemical Society Transactions*. 2021, 49(1), 93-105. DOI: 10.1042/BST20191219
- [189] Salminen A, Kaarniranta K, Hiltunen M, Kauppinen A. Krebs cycle dysfunction shapes epigenetic landscape of chromatin: novel insights into mitochondrial regulation of aging process. *Cellular Signalling*. 2014, 26(7), 1598-603. DOI: 10.1016/j.cellsig.2014.03.030
- [190] Carey BW, Finley LW, Cross JR, Allis CD, Thompson CB. Intracellular α -ketoglutarate maintains the pluripotency of embryonic stem cells. *Nature*. 2015, 518(7539), 413-416. DOI: 10.1038/nature13981
- [191] Zhang D, Tang ZY, Huang H, Zhou GL, Cui C, Weng YJ, et al. Metabolic regulation of gene expression by histone lactylation. *Nature*. 2019, 574(7779), 575-580. DOI: 10.1038/s41586-019-1678-1
- [192] Zhang Y, Song H, Li ML, Lu PR. Histone lactylation bridges metabolic reprogramming and epigenetic rewiring in driving carcinogenesis: Oncometabolite fuels oncogenic transcription. *Clinical and Translational Medicine*. 2024, 14(3), e1614. DOI: 10.1002/ctm2.1614
- [193] Yang ZJ, Zheng YQ, Gao Q. Lysine lactylation in the regulation of tumor biology. *Trends in Endocrinology and Metabolism: TEM*. 2024, 35(8), 720-731. DOI: 10.1016/j.tem.2024.01.011
- [194] Zhao P, Qiao CZ, Wang JW, Zhou Y, Zhang CH. Histone lactylation facilitates hepatocellular carcinoma progression by upregulating endothelial cell-specific molecule 1 expression. *Molecular Carcinogenesis*. 2024, 63(11), 2078-2089. DOI: 10.1002/mc.23794
- [195] Zhang C, Zhou LJ, Zhang MY, Du Y, Li C, Ren HJ, et al. H3K18 Lactylation Potentiates Immune Escape of Non-Small Cell Lung Cancer. *Cancer Research*. 2024, 84(21), 3589-3601. DOI: 10.1158/0008-5472.CAN-23-3513
- [196] Yang H, Zou XM, Yang SF, Zhang AG, Li NN, Ma Z. Identification of lactylation related model to predict prognostic, tumor infiltrating immunocytes and response of immunotherapy in gastric cancer. *Frontiers in Immunology*. 2023, 14, 1149989. DOI: 10.3389/fimmu.2023.1149989
- [197] Wang X, Liu XH, Xiao RL, Fang Y, Zhou FH, Gu MZ, et al. Histone lactylation dynamics: Unlocking the triad of metabolism, epigenetics, and immune regulation in metastatic cascade of pancreatic cancer. *Cancer Letters*. 2024, 598, 217117. DOI: 10.1016/j.canlet.2024.217117
- [198] Wang RJ, Li CW, Cheng ZY, Li MY, Shi JB, Zhang ZY, et al. H3K9 lactylation in malignant cells facilitates CD8+ T cell dysfunction and poor immunotherapy response. *Cell Reports*. 2024, 43(9), 114686. DOI: 10.1016/j.celrep.2024.114686
- [199] Anderson NM, Simon MC. The tumor microenvironment. *Current Biology: CB*. 2020, 30(16), R921-R925. DOI: 10.1016/j.cub.2020.06.081
- [200] Bharadwaj D, Mandal M. Tumor microenvironment: A playground for cells from multiple diverse origins. *Biochimica et Biophysica Acta. Reviews on Cancer*. 2024, 1879(5), 189158. DOI: 10.1016/j.bbcan.2024.189158
- [201] Ingravallo G, Tamma R, Opinto G, Annese T, Gaudio F, Specchia G, et al. The Effect of the Tumor Microenvironment on Lymphoid Neoplasms Derived from B Cells. *Diagnostics (Basel)*. 2022, 12(3), 573. DOI: 10.3390/diagnostics12030573
- [202] Wang Q, Yin XZ, Huang XT, Zhang L, Lu HJ. Impact of mitochondrial dysfunction on the antitumor effects of immune cells. *Frontiers in Immunology*. 2024, 15, 1428596. DOI: 10.3389/fimmu.2024.1428596
- [203] Porporato PE, Filigheddu N, Pedro JMB, Kroemer G, Galluzzi L. Mitochondrial metabolism and cancer. *Cell Research*. 2018, 28(3), 265-280. DOI: 10.1038/cr.2017.155
- [204] Siska PJ, Singer K, Evert K, Renner K, Kreutz M. The immunological Warburg effect: Can a metabolic-tumor-stroma score (MeTS) guide cancer immunotherapy? *Immunological Reviews*. 2020, 295(1), 187-202. DOI: 10.1111/imr.12846
- [205] Jiang MQ, Wang YC, Zhao XY, Yu JM. From metabolic byproduct to immune modulator: the role of lactate in tumor immune escape. *Frontiers in Immunology*. 2024, 15, 1492050. DOI: 10.3389/fimmu.2024.1492050
- [206] Romero-Garcia S, Moreno-Altamirano MM, Prado-Garcia H, Sánchez-García FJ. Lactate Contribution to the Tumor Microenvironment: Mechanisms, Effects on Immune Cells and Therapeutic Relevance. *Frontiers in Immunology*. 2016, 7, 52. DOI: 10.3389/fimmu.2016.00052
- [207] Ramapriyan R, Caetano MS, Barsoumian HB, Mafra ACP, Zambalde EP, Menon H, et al. Altered cancer metabolism in mechanisms of immunotherapy resistance. *Pharmacology & Therapeutics*. 2019, 195, 162-171. DOI: 10.1016/j.pharmthera.2018.11.004
- [208] Lim S, Phillips JB, Madeira da Silva L, Zhou M, Fodstad O, Owen LB, et al. Interplay between Immune Checkpoint Proteins and Cellular Metabolism. *Cancer Research*. 2017, 77(6), 1245-1249. DOI: 10.1158/0008-5472.CAN-16-1647
- [209] Xia L, Oyang L, Lin J, Tan S, Han Y, Wu N, et al. The cancer metabolic reprogramming and immune response. *Molecular Cancer*. 2021, 20(1), 28. DOI: 10.1186/s12943-021-01316-8
- [210] Song KH, Kim JH, Lee YH, Bae HC, Lee HJ, Woo SR, et al. Mitochondrial reprogramming via ATP5H loss promotes multimodal cancer therapy resistance. *The Journal of Clinical Investigation*. 2018, 128(9), 4098-4114. DOI: 10.1172/JCI96804
- [211] Ding XC, Wang LL, Zhang XD, Xu JL, Li PF, Liang H, et al. The relationship between expression of PD-L1 and HIF-1 α in glioma cells under hypoxia. *Journal of Hematology & Oncology*. 2021, 14(1), 92. DOI: 10.1186/s13045-021-01102-5
- [212] Chen F, Xue Y, Zhang W, Zhou H, Zhou Z, Chen T, et al. The role of mitochondria in tumor metastasis and advances in mitochondria-targeted cancer therapy. *Cancer Metastasis Reviews*. 2024, 43(4), 1419-1443. DOI: 10.1007/s10555-024-10211-9
- [213] Brestoff JR, Singh KK, Aquilano K, Becker LB, Berridge

- MV, Boilard E, et al. Recommendations for mitochondria transfer and transplantation nomenclature and characterization. *Nature Metabolism*. 2025, 7(1), 53-67. DOI: 10.1038/s42255-024-01200-x
- [214] Herst PM, Grasso C, Berridge MV. Metabolic reprogramming of mitochondrial respiration in metastatic cancer. *Cancer Metastasis Reviews*. 2018, 37(4), 643-653. DOI: 10.1007/s10555-018-9769-2
- [215] Bjerring JS, Khodour Y, Peterson EA, Sachs PC, Bruno RD. Intercellular mitochondrial transfer contributes to microenvironmental redirection of cancer cell fate. *The FEBS Journal*. 2025. DOI: 10.1111/febs.70002
- [216] Kuo FC, Tsai HY, Cheng BL, Tsai KJ, Chen PC, Huang YB, et al. Endothelial Mitochondria Transfer to Melanoma Induces M2-Type Macrophage Polarization and Promotes Tumor Growth by the Nrf2/HO-1-Mediated Pathway. *International Journal of Molecular Sciences*. 2024, 25(3), 1857. DOI: 10.3390/ijms25031857
- [217] Watson DC, Bayik D, Storevik S, Moreino SS, Sprowls SA, Han J, et al. GAP43-dependent mitochondria transfer from astrocytes enhances glioblastoma tumorigenicity. *Nature Cancer*. 2023, 4(5), 648-664. DOI: 10.1038/s43018-023-00556-5
- [218] Moschoi R, Imbert V, Nebout M, Chiche J, Mary D, Prebet T, et al. Protective mitochondrial transfer from bone marrow stromal cells to acute myeloid leukemic cells during chemotherapy. *Blood*. 2016, 128(2), 253-64. DOI: 10.1182/blood-2015-07-655860
- [219] Sun C, Liu X, Wang B, Wang Z, Liu Y, Di C, et al. Endocytosis-mediated mitochondrial transplantation: Transferring normal human astrocytic mitochondria into glioma cells rescues aerobic respiration and enhances radiosensitivity. *Theranostics*. 2019, 9(12), 3595-3607. DOI: 10.7150/thno.33100
- [220] Ding P, Gao C, Zhou J, Mei J, Li G, Liu D, et al. Mitochondria from osteolineage cells regulate myeloid cell-mediated bone resorption. *Nature Communications*. 2024, 15(1), 5094. DOI: 10.1038/s41467-024-49159-3
- [221] Zhang H, Yu X, Ye J, Li H, Hu J, Tan Y, et al. Systematic investigation of mitochondrial transfer between cancer cells and T cells at single-cell resolution. *Cancer Cell*. 2023, 41(10), 1788-1802.e10. DOI: 10.1016/j.ccell.2023.09.003
- [222] Cai W, Zhang J, Yu Y, Ni Y, Wei Y, Cheng Y, et al. Mitochondrial Transfer Regulates Cell Fate Through Metabolic Remodeling in Osteoporosis. *Advanced Science (Weinheim, Baden-Wurtemberg, Germany)*. 2023, 10(4), e2204871. DOI: 10.1002/advs.202204871
- [223] Baldwin JG, Heuser-Loy C, Saha T, Schelker RC, Slavkovic-Lukic D, Strieder N, et al. Intercellular nanotube-mediated mitochondrial transfer enhances T cell metabolic fitness and antitumor efficacy. *Cell*. 2024, 187(23), 6614-6630.e21. DOI: 10.1016/j.cell.2024.08.029
- [224] Headley CA, Gautam S, Olmo-Fontanez A, Garcia-Vilanova A, Dwivedi V, Akhter A, et al. Extracellular Delivery of Functional Mitochondria Rescues the Dysfunction of CD4+ T Cells in Aging. *Advanced Science (Weinheim, Baden-Wurtemberg, Germany)*. 2024, 11(5), e2303664. DOI: 10.1002/advs.202303664
- [225] Kidwell CU, Casalini JR, Pradeep S, Scherer SD, Greiner D, Bayik D, et al. Transferred mitochondria accumulate reactive oxygen species, promoting proliferation. *Elife*. 2023, 12, e85494. DOI: 10.7554/eLife.85494
- [226] Sasaki R, Luo Y, Kishi S, Ogata R, Nishiguchi Y, Sasaki T, et al. Oxidative High Mobility Group Box-1 Accelerates Mitochondrial Transfer from Mesenchymal Stem Cells to Colorectal Cancer Cells Providing Cancer Cell Stemness. *International Journal of Molecular Sciences*. 2025, 26(3), 1192. DOI: 10.3390/ijms26031192
- [227] Chang JC, Chang HS, Wu YC, Cheng WL, Lin TT, Chang HJ, et al. Mitochondrial transplantation regulates antitumor activity, chemoresistance and mitochondrial dynamics in breast cancer. *Journal of Experimental & Clinical Cancer Research: CR*. 2019, 38(1), 30. DOI: 10.1186/s13046-019-1028-z
- [228] Marlein CR, Piddock RE, Mistry JJ, Zaitseva L, Hellmich C, Horton RH, et al. CD38-Driven Mitochondrial Trafficking Promotes Bioenergetic Plasticity in Multiple Myeloma. *Cancer Research*. 2019, 79(9), 2285-2297. DOI: 10.1158/0008-5472.CAN-18-0773
- [229] Goliwas KF, Libring S, Berestesky E, Gholizadeh S, Schwager SC, Frost AR, et al. Mitochondrial transfer from cancer-associated fibroblasts increases migration in aggressive breast cancer. *Journal of Cell Science*. 2023, 136(14), jcs260419. DOI: 10.1242/jcs.260419
- [230] He X, Zhong L, Wang N, Zhao B, Wang Y, Wu X, et al. Gastric Cancer Actively Remodels Mechanical Microenvironment to Promote Chemotherapy Resistance via MSCs-Mediated Mitochondrial Transfer. *Advanced Science (Weinheim, Baden-Wurtemberg, Germany)*. 2024, 11(47), e2404994. DOI: 10.1002/advs.202404994
- [231] Qiao X, Huang N, Meng W, Liu Y, Li J, Li C, et al. Beyond mitochondrial transfer, cell fusion rescues metabolic dysfunction and boosts malignancy in adenoid cystic carcinoma. *Cell Reports*. 2024, 43(9), 114652. DOI: 10.1016/j.celrep.2024.114652
- [232] Alves AP, Mamede AC, Alves MG, Oliveira PF, Rocha SM, Botelho MF, et al. Glycolysis Inhibition as a Strategy for Hepatocellular Carcinoma Treatment? *Current Cancer Drug Targets*. 2019, 19(1), 26-40. DOI: 10.2174/1568009618666180430144441
- [233] Zheng M, Wu C, Yang K, Yang Y, Liu Y, Gao S, et al. Novel selective hexokinase 2 inhibitor Benitrobenzamide blocks cancer cells growth by targeting glycolysis. *Pharmacological Research*. 2021, 164, 105367. DOI: 10.1016/j.phrs.2020.105367
- [234] Zhao L, Kang M, Liu X, Wang Z, Wang Y, Chen H, et al. UBR7 inhibits HCC tumorigenesis by targeting Keap1/Nrf2/Bach1/HK2 and glycolysis. *Journal of Experimental & Clinical Cancer Research: CR*. 2022, 41(1), 330. DOI: 10.1186/s13046-022-02528-6
- [235] Sheng H, Tang W. Glycolysis Inhibitors for Anticancer Therapy: A Review of Recent Patents. *Recent Patents on Anti-Cancer Drug Discovery*. 2016, 11(3), 297-308. DOI: 10.2174/1574892811666160415160104
- [236] Shankar Babu M, Mahanta S, Lakhter AJ, Hato T, Paul S, Naidu SR. Lapachol inhibits glycolysis in cancer cells by targeting pyruvate kinase M2. *PLoS One*. 2018, 13(2), e0191419. DOI: 10.1371/journal.pone.0191419
- [237] Yeung C, Gibson AE, Issaq SH, Oshima N, Baumgart JT, Edessa LD, et al. Targeting Glycolysis through Inhibition of Lactate Dehydrogenase Impairs Tumor Growth in Preclinical Models of Ewing Sarcoma. *Cancer Research*. 2019, 79(19), 5060-5073. DOI: 10.1158/0008-5472.CAN-19-0217
- [238] Zhang W, Zhang SL, Hu X, Tam KY. Targeting Tumor Metabolism for Cancer Treatment: Is Pyruvate Dehydrogenase Kinases (PDKs) a Viable Anticancer Target? *International Journal of Biological Sciences*. 2015, 11(12), 1390-400. DOI: 10.7150/ijbs.13325
- [239] Bodmer M, Becker C, Meier C, Jick SS, Meier CR. Use of metformin and the risk of ovarian cancer: a case-control analysis. *Gynecologic Oncology*. 2011, 123(2), 200-4. DOI: 10.1016/j.ygyno.2011.06.038
- [240] Molina JR, Sun Y, Protopopova M, Gera S, Bandi M, Bristow C, et al. An inhibitor of oxidative phosphorylation exploits cancer vulnerability. *Nature Medicine*. 2018, 24(7), 1036-1046. DOI: 10.1038/s41591-018-0052-4
- [241] Ashton TM, McKenna WG, Kunz-Schughart LA,

- Higgins GS. Oxidative Phosphorylation as an Emerging Target in Cancer Therapy. *Clinical Cancer Research*. 2018, 24(11), 2482-2490. DOI: 10.1158/1078-0432.CCR-17-3070
- [242] Guardado Rivas MO, Stuart SD, Thach D, Dahan M, Shorr R, Zachar Z, et al. Evidence for a novel, effective approach to targeting carcinoma catabolism exploiting the first-in-class, anti-cancer mitochondrial drug, CPI-613. *PLoS One*. 2022, 17(6), e0269620. DOI: 10.1371/journal.pone.0269620
- [243] Winter M, Nait Eldjoudi A, Guette C, Hondermarck H, Bourette RP, Fovez Q, et al. Mitochondrial adaptation decreases drug sensitivity of persistent triple negative breast cancer cells surviving combinatory and sequential chemotherapy. *Neoplasia*. 2023, 46, 100949. DOI: 10.1016/j.neo.2023.100949
- [244] Casas-Benito A, Martínez-Herrero S, Martínez A. Succinate-Directed Approaches for Warburg Effect-Targeted Cancer Management, an Alternative to Current Treatments? *Cancers (Basel)*. 2023, 15(10), 2862. DOI: 10.3390/cancers15102862
- [245] Chang Y, Du R, Xia F, Xu X, Wang H, Chen X. Dysregulation of Fatty Acid Metabolism in Breast Cancer and Its Targeted Therapy. *Breast Cancer (Dove Med Press)*. 2024, 16, 825-844. DOI: 10.2147/BCTT.S496322
- [246] Komza M, Chipuk JE. Mitochondrial metabolism: A moving target in hepatocellular carcinoma therapy. *Journal of Cellular Physiology*. 2025, 240(1), e31441. DOI: 10.1002/jcp.31441
- [247] Jones SF, Infante JR. Molecular Pathways: Fatty Acid Synthase. *Clinical Cancer Research*. 2015, 21(24), 5434-8. DOI: 10.1158/1078-0432.CCR-15-0126
- [248] Marocchi F, Palluzzi F, Nicoli P, Melixetian M, Lovati G, Bertalot G, et al. Actionable Genetic Screens Unveil Targeting of AURKA, MEK, and Fatty Acid Metabolism as an Alternative Therapeutic Approach for Advanced Melanoma. *The Journal of Investigative Dermatology*. 2023, 143(10), 1993-2006.e10. DOI: 10.1016/j.jid.2023.03.1665
- [249] Iwamoto H, Abe M, Yang Y, Cui D, Seki T, Nakamura M, et al. Cancer Lipid Metabolism Confers Antiangiogenic Drug Resistance. *Cell Metabolism*. 2018, 28(1), 104-117.e5. DOI: 10.1016/j.cmet.2018.05.005
- [250] Amoedo ND, Sarlak S, Obre E, Esteves P, Bégueret H, Kieffer Y, et al. Targeting the mitochondrial trifunctional protein restrains tumor growth in oxidative lung carcinomas. *The Journal of Clinical Investigation*. 2021, 131(1), e133081. DOI: 10.1172/JCI133081
- [251] Ashley N, Poulton J. Mitochondrial DNA is a direct target of anti-cancer anthracycline drugs. *Biochemical and Biophysical Research Communications*. 2009, 378(3), 450-5. DOI: 10.1016/j.bbrc.2008.11.059
- [252] Wisnovsky SP, Wilson JJ, Radford RJ, Pereira MP, Chan MR, Laposa RR, et al. Targeting mitochondrial DNA with a platinum-based anticancer agent. *Chemistry & Biology*. 2013, 20(11), 1323-8. DOI: 10.1016/j.chembiol.2013.08.010
- [253] de la Loza MC, Wellinger RE. A novel approach for organelle-specific DNA damage targeting reveals different susceptibility of mitochondrial DNA to the anticancer drugs camptothecin and topotecan. *Nucleic Acids Research*. 2009, 37(4), e26. DOI: 10.1093/nar/gkn1087
- [254] Somuncu B, Ekmekcioglu A, Antmen FM, Ertuzun T, Deniz E, Keskin N, et al. Targeting mitochondrial DNA polymerase gamma for selective inhibition of MLH1 deficient colon cancer growth. *PLoS One*. 2022, 17(6), e0268391. DOI: 10.1371/journal.pone.0268391
- [255] Battogtokh G, Choi YS, Kang DS, Park SJ, Shim MS, Huh KM, et al. Mitochondria-targeting drug conjugates for cytotoxic, anti-oxidizing and sensing purposes: current strategies and future perspectives. *Acta Pharmaceutica Sinica. B*. 2018, 8(6), 862-880. DOI: 10.1016/j.apsb.2018.05.006
- [256] Lee JH, Kim KY, Jin H, Baek YE, Choi Y, Jung SH, et al. Self-Assembled Coumarin Nanoparticle in Aqueous Solution as Selective Mitochondrial-Targeting Drug Delivery System. *ACS Applied Materials & Interfaces*. 2018, 10(4), 3380-3391. DOI: 10.1021/acsami.7b17711
- [257] Ehsan S, Covarrubias-Zambrano O, Bossmann SH. Mitochondrial Targeting Peptide-based Nanodelivery for Cancer Treatment. *Current Protein & Peptide Science*. 2022, 23(10), 657-671. DOI: 10.2174/1389203723666220520160435
- [258] Wang Z, Guo W, Kuang X, Hou S, Liu H. Nanopreparations for mitochondria targeting drug delivery system: Current strategies and future prospective. *Asian Journal of Pharmaceutical Sciences*. 2017, 12(6), 498-508. DOI: 10.1016/j.ajps.2017.05.006
- [259] Sun Y, Yang Q, Xia X, Li X, Ruan W, Zheng M, et al. Polymeric Nanoparticles for Mitochondria Targeting Mediated Robust Cancer Therapy. *Frontiers in Bioengineering and Biotechnology*. 2021, 9, 755727. DOI: 10.3389/fbioe.2021.755727
- [260] Yamada Y, Munechika R, Satrialdi, Kubota F, Sato Y, Sakurai Y, et al. Mitochondrial Delivery of an Anticancer Drug Via Systemic Administration Using a Mitochondrial Delivery System That Inhibits the Growth of Drug-Resistant Cancer Engrafted on Mice. *Journal of Pharmaceutical Sciences*. 2020, 109(8), 2493-2500. DOI: 10.1016/j.xphs.2020.04.020
- [261] Shao X, Zhao X, Wang B, Fan J, Wang J, An H. Tumor microenvironment targeted nano-drug delivery systems for multidrug resistant tumor therapy. *Theranostics*. 2025, 15(5), 1689-1714. DOI: 10.7150/thno.103636
- [262] Ghazizadeh Y, Sharifi-Ardani SE, Tajik N, Mirzaei R, Pourahmad J. Exploring the Potential of Mitochondria-Targeted Drug Delivery for Enhanced Breast Cancer Therapy. *International Journal of Breast Cancer*. 2025, 2025, 3013009. DOI: 10.1155/ijbc/3013009
- [263] Wang X, Liu J, Durga L, Beeraka NM, Zhou R, Lu P, et al. Recent Updates on the Efficacy of Mitocans in Photo/Radio-therapy for Targeting Metabolism in Chemo/Radio-resistant Cancers: Nanotherapeutics. *Current Medicinal Chemistry*. 2025, 32(11), 2156-2182. DOI: 10.2174/0109298673259347231019121757